

PATENT COOPERATION TREATY

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371 P

PCT

NOTIFICATION OF ELECTION

(PCT Rule 61.2)

From the INTERNATIONAL BUREAU

To:

Assistant Commissioner for Patents
United States Patent and Trademark
Office
Box PCT
Washington, D.C.20231
ETATS-UNIS D'AMERIQUE

in its capacity as elected Office

Date of mailing (day/month/year)

12 May 2000 (12.05.00)

International application No.

PCT/CA99/00852

Applicant's or agent's file reference

CG/12326.19

International filing date (day/month/year)

15 September 1999 (15.09.99)

Priority date (day/month/year)

15 September 1998 (15.09.98)

Applicant

ROUSSEAU, François

1. The designated Office is hereby notified of its election made:



in the demand filed with the International Preliminary Examining Authority on:

06 April 2000 (06.04.00)



in a notice effecting later election filed with the International Bureau on:

2. The election ☒ was

was not

made before the expiration of 19 months from the priority date or, where Rule 32 applies, within the time limit under Rule 32.2(b).

The International Bureau of WIPO
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INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No. PCT/CA99/00852

I. Basis of the report

1. This report has been drawn on the basis of *(substitute sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to the report since they do not contain amendments (Rules 70.16 and 70.17).):*

Description, pages:

1,3-35	as originally filed	
2,2a	with telefax of	11/12/2000

Claims, No.:

1-14	with telefax of	11/12/2000
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2. With regard to the **language**, all the elements marked above were available or furnished to this Authority in the language in which the international application was filed, unless otherwise indicated under this item.

These elements were available or furnished to this Authority in the following language: , which is:

- ☐ the language of a translation furnished for the purposes of the international search (under Rule 23.1(b)).
- ☐ the language of publication of the international application (under Rule 48.3(b)).
- ☐ the language of a translation furnished for the purposes of international preliminary examination (under Rule 55.2 and/or 55.3).

3. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international preliminary examination was carried out on the basis of the sequence listing:

- ☐ contained in the international application in written form.
- ☐ filed together with the international application in computer readable form.
- ☐ furnished subsequently to this Authority in written form.
- ☐ furnished subsequently to this Authority in computer readable form.
- ☐ The statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.
- ☐ The statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished.

4. The amendments have resulted in the cancellation of:

- ☐ the description, pages:
- ☐ the claims, Nos.:
- ☐ the drawings, sheets:

**INTERNATIONAL PRELIMINARY
EXAMINATION REPORT**

International application No. PCT/CA99/00852

5. ☒ This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed (Rule 70.2(c)):
(Any replacement sheet containing such amendments must be referred to under item 1 and annexed to this report.)
see separate sheet

6. Additional observations, if necessary:

III. Non-establishment of opinion with regard to novelty, inventive step and industrial applicability

1. The questions whether the claimed invention appears to be novel, to involve an inventive step (to be non-obvious), or to be industrially applicable have not been examined in respect of:

- ☐ the entire international application.
- ☒ claims Nos. 1-8 and 13-14 partially with respect to industrial applicability.

because:

- ☒ the said international application, or the said claims Nos. 1-8 and 13-14 partially relate to the following subject matter which does not require an international preliminary examination (*specify*):
see separate sheet
- ☐ the description, claims or drawings (*indicate particular elements below*) or said claims Nos. are so unclear that no meaningful opinion could be formed (*specify*):
- ☐ the claims, or said claims Nos. are so inadequately supported by the description that no meaningful opinion could be formed.
- ☐ no international search report has been established for the said claims Nos. .
2. A meaningful international preliminary examination report cannot be carried out due to the failure of the nucleotide and/or amino acid sequence listing to comply with the standard provided for in Annex C of the Administrative Instructions:
- ☐ the written form has not been furnished or does not comply with the standard.
- ☐ the computer readable form has not been furnished or does not comply with the standard.

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

1. Statement

Novelty (N)	Yes: Claims 1-14
	No: Claims
Inventive step (IS)	Yes: Claims 1-14

**INTERNATIONAL PRELIMINARY
EXAMINATION REPORT**

International application No. PCT/CA99/00852

	No:	Claims	
Industrial applicability (IA)	Yes:	Claims	9-12, 14
	No:	Claims	

2. Citations and explanations
see separate sheet

VIII. Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:
see separate sheet

**INTERNATIONAL PRELIMINARY
EXAMINATION REPORT - SEPARATE SHEET**

International application No. PCT/CA99/00852

The arguments filed by the applicant with a letter of 8.12.2000 have been taken into account for establishing said report.

Point I:

The amendments filed with the letter dated 8.12.2000 introduce subject-matter which extends beyond the content of the application as filed, contrary to Article 34(2)(b) PCT. The amendments concerned are the following:

- **claim 8:** non-cancerous cells; no basis for the general term could be found in the originally filed description (p 12, l 19-20 only disclose lymphocytes).
- **claim 9:** "tissue"
- **claim 12:** no basis could be found for the claimed method starting from step b): "assaying a function...as compared to in the absence thereof".
- **page 2a:** "**germline**" mutation; no basis could be found for mutation in this specific cell type. No implicit disclosure is present for the following reasons. A predisposition for breast cancer can be created for instance due to a spontaneous mutation of DNA in other cells than germline cells. The IPEA furthermore believes that such a predisposition can well be diagnosed from somatic cells and does not need to be examined in germline cells only.

Point III:

Claims 1-8, as well as **claim 13** as long as they depend from any one of claims 1-8, relate to subject-matter considered by this Authority to be covered by the provisions of Rule 67.1(iv) PCT. Consequently, no opinion will be formulated with respect to the industrial applicability of the subject-matter of these claims (Article 34(4)(a)(i) PCT).

Point V:

Reference is made to the following documents:

- D1: AMERICAN JOURNAL OF HUMAN GENETICS,
vol. 61, no. 4 suppl, 28.10.1997 - 1.11.1997, page A64

D2: WO97/17469

1. Articles 33(2) and (3) PCT

The subject-matter of **claims 1-14** is **novel** (Article 33(2) PCT) in the light of the closest prior art D1, since it is distinguished therefrom in that the androgen receptor is used for determining an individual's predisposition of breast cancer or for screening and selecting an agent which modulates said predisposition.

Furthermore, D1 (l 11 from the bottom) only suggests a correlation of CAG repeat length of AR and breast cancer.

D2 reveals a method of predicting the risk of prostate cancer morbidity and mortality comprising determining the length of the CAG repeat of the androgen receptor gene (abstract, claim 1).

The teaching of said document can not be combined with D1 for the following reasons: a particular marker for a particular cancer (prostate (D1) or breast (D2) cancer) can not be directly transposed to a different type of cancer due to the complexity of the genetic regulation which operates at different hormonal receptors and the intricate interactions of different hormones which can differently affect, in tissue specific fashion, the transactivation of genes they regulate.

Thus, the subject-matter of **claims 1-14** is also **inventive** according to Article 33(3) PCT.

3. Industrial Applicability

For the assessment of the present **claims 1-8**, as well as **claim 13** as long as they depend from any one of claims 1-8, on the question whether they are industrially applicable, no unified criteria exist in the PCT contracting States. The patentability can also be dependent upon the formulation of the claims. The EPO, for example, does not recognize as industrially applicable the subject-matter of claims to the use of a compound in medical treatment, but may allow, however, claims to a known compound for first use in medical treatment and the use of such a compound for the manufacture

of a medicament for a new medical treatment.

Point VIII:

1. The vague and imprecise statement "spirit ... of the subject invention" (page 34, line 12) implies that the subject-matter for which protection is sought may be different to that defined in the claims, thereby resulting in lack of clarity of the claims (Article 6 PCT) when used to interpret them (see the Guidelines, C-III, 4.3a). The above statement has not been deleted to remove this defect.
2. The wording of claim 12 is unclear (Article 6 PCT), since it cannot be derived which kind of function ("a function") of the allele is assayed.
3. The broad terms "variant", "equivalent" and "mutation" used in claims 1 and 12 are vague and unclear and leave the reader in doubt as to the meaning of the technical features to which they refer, thereby rendering the definition of the subject-matter of said claim unclear (Article 6 PCT).
Furthermore, the definitions given in the description (page 21, lines 1-17, lines 18-20 and page 22, lines 3-10) do not clarify the terms, since it is not said to what extent these substances are allowed to differ from the androgen receptor gene. The terms "variant" and "mutation" are even so broad that they encompass any nucleic acid molecule, since they are not even limited by the function/s of the androgen receptor gene. Moreover, the definition of the term "equivalent" is as vague since it is silent about how many and which function/s have to remain compared to the androgen receptor gene.

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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(21) International Application Number: PCT/CA99/00852 (22) International Filing Date: 15 September 1999 (15.09.99) (30) Priority Data: 60/100,311 15 September 1998 (15.09.98) US (71) Applicant (for all designated States except US): SIGNALGENE INC. [CA/CA]; 8475 Christophe Colomb Avenue, Montreal, Quebec H2M 2N9 (CA). (72) Inventor; and (75) Inventor/Applicant (for US only): ROUSSEAU, François [CA/CA]; 2540 Lalonde, Sainte-Foy, Quebec G1W 1M7 (CA). (74) Agents: DUBUC, Jean, H. et al.; Goudreau Gage Dubuc & Martineau Walker, The Stock Exchange Tower, Suite 3400, 800 Place Victoria, P.O. Box 242, Montreal, Quebec H4Z 1E9 (CA).		(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>Without international search report and to be republished upon receipt of that report.</i>
(54) Title: MARKER AT THE ANDROGEN RECEPTOR GENE FOR DETERMINING BREAST CANCER SUSCEPTIBILITY		
(57) Abstract <p>The present invention relates to a method of determining an individual's predisposition to breast cancer, development of breast cancer, protection against breast cancer and/or responsiveness to therapy for breast cancer. The method comprises the step of determining the androgen receptor genotype at the CAG repeat locus of an individual, or at a locus in linkage disequilibrium with the CAG repeat locus, thereby determining an individual's predisposition to breast cancer, development of breast cancer, protection against breast cancer and/or responsiveness to therapy for breast cancer.</p>		

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5 Breast cancers have the same clinical characteristics in older as in younger women. Cancer is usually suspected when changes are noted on mammography or when a breast lesion is seen or felt. Lesions usually can be felt as firm nodules within the breast. Ulcerations may occur, and lesions within or near the nipple may produce discharge. Sometimes breast cancer is discovered only after metastatic lesions cause bone fractures, neurologic changes, hypercalcemia, liver failure, or ascites.

10 When a tumor is detected by physical examination, bilateral mammograms are normally obtained to rule out occult lesions. Certain radiographic images, such as speckled calcifications or tissue infiltration, suggest cancer, while a cystic appearance suggests a benign process. Even an apparently benign finding on mammogram requires further evaluation. Generally the diagnosis is established by fine needle aspiration. Fine needle aspiration allows collection and cytological examination of cystic fluid and is helpful in planning definitive treatment of breast cancer. Although a positive result on fine needle aspiration is diagnostic, a negative result is usually followed by an open biopsy. Now a day, there is still no specific test for assaying predisposition or resistance to breast cancer.

20 Since the discovery of the human androgen receptor (AR) gene, mutations in this gene have been associated with Kennedy's disease (spinobulbarmuscular atrophy), with various degrees of androgen insensitivity and with prostate cancer. Thus, an association between the AR gene and breast cancer has yet to be reported.

25 There thus remains a need to provide a genetic assay for determining the predisposition and/or resistance to breast cancer, development of breast cancer and responsiveness to therapeutic modalities.

While some markers have been identified as genetic determinants for breast cancer and/or as risk factors to develop same (i.e. BRCA1 and BRCA2), there remains a need to identify new markers therefor.

30 More specifically, there remains a need to provide means to determine a

predisposition to breast cancer and/or responsiveness to therapy to breast cancer, by analyzing allelic variations in genes associated with breast cancer. In addition there remains a need to identify patients who are likely to benefit from a particular prevention or therapeutic treatment program. Further, there remains a need to provide assays to screen for compounds (i.e. hormones, molecules acting on hormone receptors or other agents) that could be beneficial to patients.

The present invention seeks to meet these and other needs. The present description refers to a number of documents, the content of which is herein incorporated by reference, in their entirety.

SUMMARY OF THE INVENTION

One aim of the present invention is to provide a genetic assay for determining the predisposition to breast cancer and/or response to breast cancer treatment.

Another aim of the present invention is to use a polymorphism of the androgen receptor (AR) gene or an equivalent thereof as a marker for breast cancer susceptibility and/or response to breast cancer preventive or curative therapy. A polymorphism of the androgen receptor (AR) gene, or any polymorphism in linkage disequilibrium therewith, can be used as a test for breast cancer susceptibility, for responsiveness to treatment of breast cancer, for breast cancer prognosis or severity, or as a means to classify patients in clinical trial for breast cancer (screening, diagnosis, prognosis or treatment).

One of a polymorphism of the AR gene, or any polymorphism in linkage disequilibrium therewith, can further be used as a test for screening drugs for breast cancer or for determining the best treatment therefor.

Broadly, the present invention aims at providing a method of determining the length of a CAG repeat polymorphism in the androgen receptor gene, wherein this determination can be correlated with a predisposition or a

protection to breast cancer. This determination can be based on a variety of genotyping methods at the DNA, RNA or protein level.

Another aim of the present invention is to provide a method of prognosing and/or forecasting the development of breast cancer in a patient, which comprises determining a CAG-repeat polymorphism of the AR gene, or
5 any polymorphism in linkage disequilibrium therewith, in a biological sample of the patient, wherein a determination of the length of the CAG repeat shows a significant association with breast cancer.

In a particular embodiment, the determination of the
10 polymorphism at the CAG repeat of the AR gene enables to show that the shortest alleles or a combination of the shortest alleles are associated with the smallest breast cancer risk and the mid to long alleles or a combination of the intermediate and longest alleles are associated with the highest breast cancer risk (a combination of the longest alleles is associated with the highest risks of
15 breast cancer). Of importance, the variations of polymorphisms at the CAG repeat locus of AR (or of an equivalent or marker in linkage disequilibrium therewith) can account for a significant proportion of all cases of breast cancer. Indeed, the number of breast cancer cases attributable to a variation at this AR locus is at least three times greater than that attributable to the BRCA1 and
20 BRCA2 genes.

The present invention also relates to vectors, including expression vectors harboring an AR gene (or fragment or fusion thereof) having a genotype in accordance with the present invention (i.e. a predisposing genotype, long CAG repeats, or alternatively, a protecting genotype, short CAG
25 repeats; or other genotypes isolated from patients or genetically engineered), cells harboring such vectors, and non-human animals harboring such vectors or cells.

Another aim of the present invention is to provide means of identifying young women that will be at risk of developing breast cancer and to
30 categorize those that are likely to respond significantly to preventive therapy.

An aim of the present invention is thus to provide means of identification of target sub-groups of women for breast cancer prevention measures/programs.

Another aim of the present invention is to provide means to determine which sub-group of women will most benefit from breast cancer treatment(s) and eventually predict their response to therapy or choose the optimal preventive pharmacotherapy.

Another aim of the present invention is to identify means of predicting and managing interventions for breast cancer as well as identifying and/or characterizing biological parameters which could enable the establishment of population-based breast cancer prevention and intervention programs.

In addition, it is an aim of the present invention to provide a method of selecting alleles of the AR gene or in linkage disequilibrium therewith, which is suitable for designing an assay to screen compounds which can modulate the activity of an androgen receptor.

Another aim of the present invention is to provide an assay to screen for drugs for the treatment and/or prevention of breast cancer. Having identified alleles which predispose to breast cancer (and those which predispose to a "resistance" to breast cancer), assays can be set-up to screen agents and select drugs which could be used in the treatment or prevention of breast cancer. Since some alleles of the AR have been shown to affect the functionality of the androgen receptor (Tut et al. 1997, J. Clin. Endocrinol. 89(11):3777-3782), assays could be designed based on chosen genotypes of the AR gene. A non-limiting example of a type of assay which could be designed includes, *cis-trans* assays similar to those described in USP 4,981,784. For example, a *cis-trans* assay could be set-up, based on the use of a genotype of AR, shown here to predispose to breast cancer (i.e. the long CAG alleles in the AR gene) as compared to a genotype of AR, shown here to be associated with lower risk of breast cancer, and used to screen compounds. A non-limiting example of such an assay could be based on 2 cell lines (one

expressing a predisposing genotype of AR and one expressing a non-predisposing genotype of AR) which could be used in parallel to screen for AR-function modulating compounds. Of course, it will be understood that the cell line expressing the non-predisposing genotype of AR (the shorter alleles) can
5 be used as a positive control for the functionality of the androgen receptor.

It is thus an aim of the present invention to provide the means to identify compounds which could positively modulate the function of AR having a breast cancer predisposing genotype (such as the long CAG alleles), to the level of the protecting genotype thereof (such as the short CAG alleles).

10 In a particular embodiment, such assays can be designed using cells from patients having a known genotype at the loci of the present invention, these cells harboring recombinant vectors could enable an assessment of the functionality of the AR and dissect the structure-function relationship of the androgen receptor and its role in breast cancer.

15 It shall be understood that the polymorphism of the AR and/or the determination of allelic variations in the AR gene can be combined to the determination of allelic variations in other genes/markers linked to the predisposition to breast cancer and/or responsiveness to therapy therefor. This combination of genotype analyses could lead to better diagnoses programs
20 and/or treatment of breast cancer. Non-limiting examples of such markers include BRCA1 and BRCA2.

It shall also be understood that although breast cancer is significantly more preponderant in women, it can also be a deadly disease in men. Thus, the present invention is meant to also cover men.

25 In accordance with the present invention, there is therefore provided a method of determining an individual's predisposition to breast cancer, development of breast cancer and/or responsiveness to therapy for breast cancer, which comprises determining a genotype at the CAG-repeat locus of the androgen receptor (directly or indirectly by linkage disequilibrium) in a biological
30 sample of the individual and analyzing allelic variation in the androgen receptor

of the individual, thereby determining an individual's predisposition to breast cancer, development of breast cancer and/or responsiveness to therapy therefor.

5 In accordance with the present invention there is provided a method for determining susceptibility to breast cancer, and/or response to therapy therefor. The method comprises the step of determining the androgen receptor genotype of the individual, thereby determining an individual's susceptibility to breast cancer and/or response to therapy therefor.

10 Numerous methods for determining a genotype are known and available to the skilled artisan. All these genotype determination methods are within the scope of the present invention. Non-limiting examples of genotype determination include a restriction endonuclease digestion, a hybridization with allele specific oligonucleotides, a sequencing of the polymorphism, and an amplification of a segment of the androgen receptor (i.e. by PCR).

15 In accordance with the present invention, there is therefore provided a method of determining an individual's predisposition to breast cancer, development of breast cancer and/or responsiveness to therapy therefor, which comprises determining androgen receptor polymorphism (directly or indirectly using a marker in linkage disequilibrium with the CAG repeat polymorphism) in
20 a biological sample of the individual and analyzing allelic variation in the androgen receptor gene of the individual, thereby determining an individual's predisposition to breast cancer, development of breast cancer and/or responsiveness to therapy therefor.

25 In accordance with one embodiment of the invention, there is provided a specific model for use in prediction of breast cancer susceptibility and prognosis. The model comprises an androgen receptor gene polymorphisms at the CAG repeat locus, that allows to identify a subset of women that are at significantly increased risk of breast cancer as compared to those bearing other variant of this gene.

In accordance with a preferred embodiment of the present invention, a single gene, the androgen receptor gene, has been identified as such a target to assess this predisposition.

5 In accordance with the present invention, the androgen receptor polymorphism, without limitation, is selected from the CAG repeats located in the first exon of the AR gene, or any DNA variant or mutation which shows some degree of linkage disequilibrium with one of the polymorphisms at the CAG-repeat locus of the AR gene.

10 In some embodiments, the method of the present invention includes detecting the androgen receptor polymorphism by analyzing the restriction fragment length polymorphisms using an endonuclease digestion. The method can further include a step prior to the androgen receptor gene digestion, wherein at least a fragment of the androgen receptor is amplified, for example, by polymerase chain reaction.

15 In accordance with a preferred embodiment of the present invention, a pair of primers is designed to specifically amplify a segment of the androgen receptor. In an especially preferred embodiment, the region of the AR gene which is amplified is in exon 1. This pair of primers is preferably derived from a nucleic acid sequence of the androgen receptor gene or flanking portion thereof, to amplify a segment of the androgen receptor gene, as commonly
20 known. Of course, other primer pairs can be designed, based on the known sequence of the AR gene. Method to design primer pairs from known sequences are commonly known in the art.

25 In accordance with a preferred embodiment of the present invention, primers used for amplifying the segment of the androgen receptor are defined as:

5'-TCCAGAATCTGTTCCAGAGCGTGC-3 (SEQ ID NO:1); and

5'-GCTGTGAAGGTTGCTGTTCTCAT-3' (SEQ ID NO:2).

The polymorphism of the androgen receptor gene can be detected using at least one oligonucleotide specific to the normal or variant androgen receptor gene allele.

5 The present invention also provides a kit for determining predisposition to low, intermediate or high risk of breast cancer of a patient, which includes at least a probe specific for the androgen receptor; a polymorphism selected from the group consisting of a CAG repeat and other polymorphisms in linkage disequilibrium with the CAG repeat polymorphism.

10 In one embodiment, the present invention provides a specific detection of the CAG repeat polymorphism of the AR gene using a nucleic acid for the specific detection of this AR polymorphism in a sample comprising the above-described CAG-repeat-containing nucleic acid sequence (i.e. DNA, RNA, cDNA) and at least a nucleic acid sequence which binds under stringent conditions to the CAG-repeat-containing nucleic acid sequence.

15 In one preferred embodiment, the present invention relates to nucleic acid probes which are complementary to a CAG-repeat-containing nucleic acid sequence, consisting of at least 10 consecutive nucleotides (preferably, 15, 20, 25, or 30) and which specifically hybridize to the AR nucleic acid sequence comprising the CAG repeat polymorphism under high stringency
20 condition.

In one embodiment of the above described method, a nucleic acid probe is immobilized on a solid support. Non-limiting examples of solid supports include plastics (i.e. polycarbonate), acrylic resins (i.e. polyacrylamide and latex beads); and carbohydrates (i.e. agarose and sepharose). Techniques
25 for coupling nucleic acid probes to solid supports are well known in the art.

Similarly to the probes of the present invention, the antibodies of the present invention can be immobilized on a solid support. As known in the art, similar supports as those used for probe immobilization can be used for antibody immobilization on a solid support. Also well known in the art are the
30 techniques for coupling antibodies to such solid supports. The immobilized

antibodies of the present invention can be used for *in vitro*, *in vivo*, and *in situ* assays as well as in immunochromatography according to known methods.

Non-limiting examples of test samples suitable for carrying the methods of the present invention include, cells or nucleic acid extracts of
5 cells, or biological fluids. Of course, the type of test sample used can vary according to the assay format, the method of detection, and the particular needs of the clinical practitioner which will readily adapt the methods of preparation of the sample and the method of detection so that they are compatible, in accordance with the knowledge in the art.

10 In accordance with one embodiment of the present invention, the allelic variation in the androgen receptor gene is analyzed indirectly using a nucleic acid variant, or equivalent in linkage disequilibrium with a CAG repeat. The allelic variation in the androgen receptor gene can also be analyzed directly by determining the number of CAG repeat within the androgen receptor gene.

15 In accordance with the present invention, the polymorphism of the androgen receptor (AR) gene can be used as a marker for breast cancer susceptibility. The polymorphism in linkage disequilibrium with the markers used can also be used as a test for breast cancer susceptibility, or for responsiveness to treatment for breast cancer, for breast cancer prognosis or severity, or as a
20 means to classify patients in clinical trials for breast cancer (screening, diagnosis, prognosis or treatment).

In order to provide a clear and consistent understanding of terms used in the present description, a number of definitions are provided hereinbelow.

25 As used herein the term "RFLP" refers to restriction fragment length polymorphism.

The terms "polymorphism", "DNA polymorphism" and the like, refer to any sequence in the human genome which exists in more than one version or variant in the population.

The term "linkage disequilibrium" refers to any degree of non-random genetic association between one or more allele(s) of two different polymorphic DNA sequences, that is due to the physical proximity of the two loci. Linkage disequilibrium is present when two DNA segments that are very close to each other on a given chromosome will tend to remain unseparated for several generations with the consequence that alleles of a DNA polymorphism (or marker) in one segment will show a non-random association with the alleles of a different DNA polymorphism (or marker) located in the other DNA segment nearby. Hence, testing of one of a marker in linkage disequilibrium with the polymorphisms of the present invention at the AR gene (indirect testing), will give almost the same information as testing for the CAG repeat polymorphism of the AR gene directly. This situation is encountered throughout all the human genome when two DNA polymorphisms that are very close to each other are studied. Such a linkage disequilibrium has been reported with several polymorphisms in several genes (i.e. the vitamin D receptor gene [Morrisson et al., 1994, Nature 367:284-287, and USP 5,593,033]). Various degrees of linkage disequilibrium can be encountered between two genetic markers so that some are more closely associated than others.

The terms "androgen receptor polymorphism" or "genetic marker" are intended to include, without limitation, the CAG-repeat polymorphism in exon 1, and any other allelic variant of the androgen receptor gene that show some degree of linkage disequilibrium in any population subgroup with at least one of the above-mentioned androgen receptor polymorphisms.

The androgen receptor gene polymorphism sites in accordance with the present invention can be located within the androgen receptor gene, or on each side thereof, provided that is on the same chromosome and in linkage disequilibrium with the AR polymorphism of the present invention. Distances between markers in linkage disequilibrium can vary widely (below 50 kb to more than 1 mega base) depending on the genetic

structure of the population and is ascertainable by a statistically significant association between the markers.

It shall be recognized by the person skilled in the art to which the present invention pertains, that since some of the polymorphisms herein identified in the AR gene can be within the coding region of the gene and therefore expressed, that the present invention should not be limited to the identification of polymorphisms at the DNA level (whether on genomic DNA, amplified DNA, cDNA or the like). Indeed, the herein-identified polymorphisms could be detected at the mRNA or protein level. Such detections of polymorphism identification on mRNA or protein are known in the art. Non-limiting examples include detection based on oligos designed to hybridize to mRNA or ligands such as antibodies which are specific to the encoded polymorphism (i.e. specific to the protein fragment encoded by the CAG repeat for example).

Since some of the polymorphisms of the present invention are expressed, one of the advantages of the present invention is to enable a determination of the polymorphisms in the AR gene, in easily obtainable cells which express these genes. A non-limiting example thereof is lymphocytes, thereby enabling a genotyping from a simple blood sample.

Nucleotide sequences are presented herein by single strand, in the 5' to 3' direction, from left to right, using the one letter nucleotide symbols as commonly used in the art and in accordance with the recommendations of the IUPAC-IUB Biochemical Nomenclature Commission.

Unless defined otherwise, the scientific and technological terms and nomenclature used herein have the same meaning as commonly understood by a person of ordinary skill to which this invention pertains. Generally, the procedures for cell cultures, infection, molecular biology methods and the like are common methods used in the art. Such standard techniques can be found in reference manuals such as for example Sambrook et al. (1989,

Molecular Cloning -A Laboratory Manual, Cold Spring Harbor Laboratories) and Ausubel et al. (1994, Current Protocols in Molecular Biology, Wiley, New York).

The present description refers to a number of routinely used recombinant DNA (rDNA) technology terms. Nevertheless, definitions of selected examples of such rDNA terms are provided for clarity and consistency.

As used herein, "nucleic acid molecule", refers to a polymer of nucleotides. Non-limiting examples thereof include DNA (i.e. genomic DNA, cDNA) and RNA molecules (i.e. mRNA). The nucleic acid molecule can be obtained by cloning techniques or synthesized. DNA can be double-stranded or single-stranded (coding strand or non-coding strand [antisense]).

The term "recombinant DNA" as known in the art refers to a DNA molecule resulting from the joining of DNA segments. This is often referred to as genetic engineering.

The term "DNA segment", is used herein, to refer to a DNA molecule comprising a linear stretch or sequence of nucleotides. This sequence when read in accordance with the genetic code, can encode a linear stretch or sequence of amino acids which can be referred to as a polypeptide, protein, protein fragment and the like.

The terminology "amplification pair" refers herein to a pair of oligonucleotides (oligos) of the present invention, which are selected to be used together in amplifying a selected nucleic acid sequence by one of a number of types of amplification processes, preferably a polymerase chain reaction. Other types of amplification processes include ligase chain reaction, strand displacement amplification, or nucleic acid sequence-based amplification, as explained in greater detail below. As commonly known in the art, the oligos are designed to bind to a complementary sequence under selected conditions.

The nucleic acid (i.e. DNA or RNA) for practicing the present invention may be obtained according to well known methods.

Oligonucleotide probes or primers of the present invention may be of any suitable length, depending on the particular assay format and the

particular needs and targeted genomes employed. In general, the oligonucleotide probes or primers are at least 12 nucleotides in length, preferably between 15 and 24 molecules, and they may be adapted to be especially suited to a chosen nucleic acid amplification system. As commonly known in the art, the oligonucleotide probes and primers can be designed by taking into consideration the melting point of hybridization thereof with its targeted sequence (see below and in Sambrook et al., 1989, Molecular Cloning - A Laboratory Manual, 2nd Edition, CSH Laboratories; Ausubel et al., 1989, in Current Protocols in Molecular Biology, John Wiley & Sons Inc., N.Y.).

The term "oligonucleotide" or "DNA" molecule or sequence refers to a molecule comprised of the deoxyribonucleotides adenine (A), guanine (G), thymine (T) and/or cytosine (C), in a double-stranded form, and comprises or includes a "regulatory element" according to the present invention, as the term is defined herein. The term "oligonucleotide" or "DNA" can be found in linear DNA molecules or fragments, viruses, plasmids, vectors, chromosomes or synthetically derived DNA. As used herein, particular double-stranded DNA sequences may be described according to the normal convention of giving only the sequence in the 5' to 3' direction.

"Nucleic acid hybridization" refers generally to the hybridization of two single-stranded nucleic acid molecules having complementary base sequences, which under appropriate conditions will form a thermodynamically favored double-stranded structure. Examples of hybridization conditions can be found in the two laboratory manuals referred above (Sambrook et al., 1989, *supra* and Ausubel et al., 1989, *supra*) and are commonly known in the art. In the case of a hybridization to a nitrocellulose filter, as for example in the well known Southern blotting procedure, a nitrocellulose filter can be incubated overnight at 65°C with a labeled probe in a solution containing 50% formamide, high salt (5 x SSC or 5 x SSPE), 5 x Denhardt's solution, 1% SDS, and 100 µg/ml denatured carrier DNA (i.e. salmon sperm DNA). The non-specifically binding probe can then be washed off the

filter by several washes in 0.2 x SSC/0.1% SDS at a temperature which is selected in view of the desired stringency: room temperature (low stringency), 42°C (moderate stringency) or 65°C (high stringency). The selected temperature is based on the melting temperature (T_m) of the DNA hybrid. Of course, RNA-DNA hybrids can also be formed and detected. In such cases, the conditions of hybridization and washing can be adapted according to well known methods by the person of ordinary skill. Stringent conditions will be preferably used (Sambrook et al., 1989, *supra*).

Probes of the invention can be utilized with naturally occurring sugar-phosphate backbones as well as modified backbones including phosphorothioates, dithionates, alkyl phosphonates and α -nucleotides and the like. Modified sugar-phosphate backbones are generally taught by Miller, 1988, Ann. Reports Med. Chem. 23:295 and Moran et al., 1987, Nucleic acid molecule. Acids Res., 14:5019. Probes of the invention can be constructed of either ribonucleic acid (RNA) or deoxyribonucleic acid (DNA), and preferably of DNA.

The types of detection methods in which probes can be used include Southern blots (DNA detection), dot or slot blots (DNA, RNA), and Northern blots (RNA detection). Although less preferred, labeled proteins could also be used to detect a particular nucleic acid sequence to which it binds. More recently, PNAs have been described (Nielsen et al. 1999, Current Opin. Biotechnol. 10:71-75). PNAs could also be used to detect the polymorphisms of the present invention. Other detection methods include kits containing probes on a dipstick setup and the like.

Although the present invention is not specifically dependent on the use of a label for the detection of a particular nucleic acid sequence, such a label might be beneficial, by increasing the sensitivity of the detection. Furthermore, it enables automation. Probes can be labeled according to numerous well known methods (Sambrook et al., 1989, *supra*). Non-limiting examples of labels include ^3H , ^{14}C , ^{32}P , and ^{35}S . Non-limiting examples of detectable markers include ligands, fluorophores, chemiluminescent agents,

enzymes, and antibodies. Other detectable markers for use with probes, which can enable an increase in sensitivity of the method of the invention, include biotin and radionucleotides. It will become evident to the person of ordinary skill that the choice of a particular label dictates the manner in which it is bound to the probe.

As commonly known, radioactive nucleotides can be incorporated into probes of the invention by several methods. Non-limiting examples thereof include kinasing the 5' ends of the probes using gamma ³²P ATP and polynucleotide kinase, using the Klenow fragment of Pol I of *E. coli* in the presence of radioactive dNTP (i.e. uniformly labeled DNA probe using random oligonucleotide primers in low-melt gels), using the SP6/T7 system to transcribe a DNA segment in the presence of one or more radioactive NTP, and the like.

As used herein, "oligonucleotides" or "oligos" define a molecule having two or more nucleotides (ribo or deoxyribonucleotides). The size of the oligo will be dictated by the particular situation and ultimately on the particular use thereof and adapted accordingly by the person of ordinary skill. An oligonucleotide can be synthesized chemically or derived by cloning according to well known methods.

As used herein, a "primer" defines an oligonucleotide which is capable of annealing to a target sequence, thereby creating a double stranded region which can serve as an initiation point for DNA synthesis under suitable conditions.

Amplification of a selected, or target, nucleic acid sequence may be carried out by a number of suitable methods. See generally Kwoh et al., 1990, *Am. Biotechnol. Lab.* 8:14-25. Numerous amplification techniques have been described and can be readily adapted to suit particular needs of a person of ordinary skill. Non-limiting examples of amplification techniques include polymerase chain reaction (PCR), ligase chain reaction (LCR), strand displacement amplification (SDA), transcription-based

amplification, the Q β replicase system and NASBA (Kwoh et al., 1989, Proc. Natl. Acad. Sci. USA 86, 1173-1177; Lizardi et al., 1988, BioTechnology 6:1197-1202; Malek et al., 1994, Methods Mol. Biol., 28:253-260; and Sambrook et al., 1989, *supra*). Preferably, amplification will be carried out using PCR.

5 Polymerase chain reaction (PCR) is carried out in accordance with known techniques. See, e.g., U.S. Pat. Nos. 4,683,195; 4,683,202; 4,800,159; and 4,965,188 (the disclosures of all three U.S. Patent are incorporated herein by reference). In general, PCR involves, a treatment of a nucleic acid sample (e.g., in the presence of a heat stable DNA polymerase)
10 under hybridizing conditions, with one oligonucleotide primer for each strand of the specific sequence to be detected. An extension product of each primer which is synthesized is complementary to each of the two nucleic acid strands, with the primers sufficiently complementary to each strand of the specific sequence to hybridize therewith. The extension product synthesized from each primer can
15 also serve as a template for further synthesis of extension products using the same primers. Following a sufficient number of rounds of synthesis of extension products, the sample is analysed to assess whether the sequence or sequences to be detected are present. Detection of the amplified sequence may be carried out by visualization following EtBr staining of the DNA following gel
20 electrophoresis, or using a detectable label in accordance with known techniques, and the like. For a review on PCR techniques (see PCR Protocols, A Guide to Methods and Amplifications, Michael et al. Eds, Acad. Press, 1990).

Ligase chain reaction (LCR) is carried out in accordance with known techniques (Weiss, 1991, Science 254:1292). Adaptation of the
25 protocol to meet the desired needs can be carried out by a person of ordinary skill. Strand displacement amplification (SDA) is also carried out in accordance with known techniques or adaptations thereof to meet the particular needs (Walker et al., 1992, Proc. Natl. Acad. Sci. USA 89:392-396; and *ibid.*, 1992, Nucleic Acids Res. 20:1691-1696).

As used herein, the term "gene" is well known in the art and relates to a nucleic acid sequence defining a single protein or polypeptide. A "structural gene" defines a DNA sequence which is transcribed into RNA and translated into a protein having a specific amino acid sequence thereby giving rise the a specific polypeptide or protein.

A "heterologous" (i.e. a heterologous gene) region of a DNA molecule is a subsegment segment of DNA within a larger segment that is not found in association therewith in nature. The term "heterologous" can be similarly used to define two polypeptidic segments not joined together in nature.

10 Non-limiting examples of heterologous genes include reporter genes such as luciferase, chloramphenicol acetyl transferase, β -galactosidase, and the like which can be juxtaposed or joined to heterologous control regions or to heterologous polypeptides.

The term "vector" is commonly known in the art and defines a plasmid DNA, phage DNA, viral DNA and the like, which can serve as a DNA vehicle into which DNA of the present invention can be cloned.

15 Numerous types of vectors exist and are well known in the art.

The term "expression" defines the process by which a gene is transcribed into mRNA (transcription), the mRNA is then being translated (translation) into one polypeptide (or protein) or more.

20

The terminology "expression vector" defines a vector or vehicle as described above but designed to enable the expression of an inserted sequence following transformation into a host. The cloned gene (inserted sequence) is usually placed under the control of control element sequences such as promoter sequences. The placing of a cloned gene under such control sequences is often referred to as being operably linked to control elements or sequences.

25

Operably linked sequences may also include two segments that are transcribed onto the same RNA transcript. Thus, two sequences, such as a promoter and a "reporter sequence" are operably linked

30

if transcription commencing in the promoter will produce an RNA transcript of the reporter sequence. In order to be "operably linked" it is not necessary that two sequences be immediately adjacent to one another.

5 Expression control sequences will vary depending on whether the vector is designed to express the operably linked gene in a prokaryotic or eukaryotic host or both (shuttle vectors) and can additionally contain transcriptional elements such as enhancer elements, termination sequences, tissue-specificity elements, and/or translational initiation and termination sites.

10 Prokaryotic expressions are useful for the preparation of large quantities of the protein encoded by the DNA sequence of interest. This protein can be purified according to standard protocols that take advantage of the intrinsic properties thereof, such as size and charge (i.e. SDS gel electrophoresis, gel filtration, centrifugation, ion exchange chromatography...).
15 In addition, the protein of interest can be purified via affinity chromatography using polyclonal or monoclonal antibodies. The purified protein can be used for therapeutic applications.

The DNA construct can be a vector comprising a promoter that is operably linked to an oligonucleotide sequence of the present invention,
20 which is in turn, operably linked to a heterologous gene, such as the gene for the luciferase reporter molecule. "Promoter" refers to a DNA regulatory region capable of binding directly or indirectly to RNA polymerase in a cell and initiating transcription of a downstream (3' direction) coding sequence. For purposes of the present invention, the promoter is bound at its 3' terminus by the
25 transcription initiation site and extends upstream (5' direction) to include the minimum number of bases or elements necessary to initiate transcription at levels detectable above background. Within the promoter will be found a transcription initiation site (conveniently defined by mapping with S1 nuclease), as well as protein binding domains (consensus sequences) responsible for the
30 binding of RNA polymerase. Eukaryotic promoters will often, but not always,

contain "TATA" boxes and "CCAT" boxes. Prokaryotic promoters contain Shine-Dalgarno sequences in addition to the -10 and -35 consensus sequences.

In accordance with one embodiment of the present invention, an expression vector can be constructed to assess the functionality of specific alleles of the AR gene and of the interaction of such alleles. Non-limiting examples of such expression vectors include a vector comprising the androgen responsive element (the cis sequences [i.e. DNA sequence to which a factor binds] enabling androgen-dependent modulating effects of promoter activity are known in the art) operably linked to a chosen promoter and modulating the activity thereof, the promoter driving the expression of a reporter gene. When such a vector is transfected in a cell expressing AR, the modulating effect of the promoter activity can be assessed by determining the level of expression of the reporter gene. In one embodiment, the vector is transfected into a cell of a patient having the genotype of AR shown herein to be associated with a low risk of breast cancer, or in a cell from a patient having the genotype of AR shown herein to be associated with a moderate or high risk of breast cancer. These cells can serve to screen for compounds that modulate the promoter activity, in order to identify compounds that could be used to treat especially, patients predicted to be at moderate or high risk of breast cancer. Of course, it will be understood that the AR gene expressed by these cells can be modified at will (i.e. by *in vitro* mutagenesis or the like). Similarly, numerous combinations of genotypes can be tested in such assays to dissect the functional relationship between the AR genotype and its function in androgen-dependent function and/or its function in breast cancer. It will also be clear to the skilled artisan, that such indicator cells expressing AR, could also be engineered by choosing a cell line and transfecting thereinto, chosen genotypes of AR and one expression vector as described above. Non-human transgenic animals expressing chosen alleles of AR could also be prepared and used to screen compounds that affect androgen receptor function and possibly overcome a predisposition to breast cancer.

As used herein, the designation "functional derivative" denotes, in the context of a functional derivative of a sequence whether an nucleic acid or amino acid sequence, a molecule that retains a biological activity (either function or structural) that is substantially similar to that of the original sequence. This functional derivative or equivalent may be a natural derivative or may be prepared synthetically. Such derivatives include amino acid sequences having substitutions, deletions, or additions of one or more amino acids, provided that the biological activity of the protein is conserved. The same applies to derivatives of nucleic acid sequences which can have substitutions, deletions, or additions of one or more nucleotides, provided that the biological activity of the sequence is generally maintained. When relating to a protein sequence, the substituting amino acid as chemico-physical properties which are similar to that of the substituted amino acid. The similar chemico-physical properties include, similarities in charge, bulkiness, hydrophobicity, hydrophylicity and the like. The term "functional derivatives" is intended to include "fragments", "segments", "variants", "analogs" or "chemical derivatives" of the subject matter of the present invention.

Thus, the term "variant" refers herein to a protein or nucleic acid molecule which is substantially similar in structure and biological activity to the protein or nucleic acid of the present invention.

The functional derivatives of the present invention can be synthesized chemically or produced through recombinant DNA technology. all these methods are well known in the art.

As used herein, "chemical derivatives" is meant to cover additional chemical moieties not normally part of the subject matter of the invention. Such moieties could affect the physico-chemical characteristic of the derivative (i.e. solubility, absorption, half life and the like, decrease of toxicity). Such moieties are exemplified in Remington's Pharmaceutical Sciences (1980). Methods of coupling these chemical-physical moieties to a polypeptide are well known in the art.

The term "allele" defines an alternative form of a gene which occupies a given locus on a chromosome.

As commonly known, a "mutation" is a detectable change in the genetic material which can be transmitted to a daughter cell. As well known, a mutation can be, for example, a detectable change in one or more deoxyribonucleotide. For example, nucleotides can be added, deleted, substituted for, inverted, or transposed to a new position. Spontaneous mutations and experimentally induced mutations exist. The result of a mutations of nucleic acid molecule is a mutant nucleic acid molecule. A mutant polypeptide can be encoded from this mutant nucleic acid molecule.

As used herein, the term "purified" refers to a molecule having been separated from a cellular component. Thus, for example, a "purified protein" has been purified to a level not found in nature. A "substantially pure" molecule is a molecule that is lacking in all other cellular components.

As used herein, the terms "molecule", "compound", or "agent" are used interchangeably and broadly to refer to natural, synthetic or semi-synthetic molecules or compounds. The term "molecule" therefore denotes for example chemicals, macromolecules, cell or tissue extracts (from plants or animals) and the like. Non limiting examples of molecules include nucleic acid molecules, peptides, ligands, including antibodies, carbohydrates and pharmaceutical agents. The agents can be selected and screened by a variety of means including random screening, rational selection and by rational design using for example protein or ligand modelling methods such as computer modelling. The terms "rationally selected" or "rationally designed" are meant to define compounds which have been chosen based on the configuration of the interaction domains of the present invention. As will be understood by the person of ordinary skill, macromolecules having non-naturally occurring modifications are also within the scope of the term "molecule". For example, peptidomimetics, well known in the pharmaceutical industry and generally referred to as peptide analogs can be generated by modelling as mentioned

above. Similarly, in a preferred embodiment, the polypeptides of the present invention are modified to enhance their stability. It should be understood that in most cases this modification should not alter the biological activity of the protein. The molecules identified in accordance with the teachings of the present invention have a therapeutic value in diseases or conditions in which a
5 apparently lower activity and/or level of the AR is linked to a genotype of AR identified in accordance with the present invention. Alternatively, the molecules identified in accordance with the teachings of the present invention find utility in the development of compounds which can modulate the activity and/or level of
10 the androgen receptor in an animal and/or overcome a predisposition to breast cancer.

As used herein, agonists and antagonists also include potentiators of known compounds with such agonist or antagonist properties. In one embodiment, modulators of the level or the activity of the AR can be
15 identified and selected by contacting the indicator cell with a compound or mixture or library of molecules for a fixed period of time. In certain embodiments, the "breast cancer-low risk-associated alleles" of the AR gene can be used as positive controls.

An indicator cell in accordance with the present invention
20 can be used to identify antagonists. For example, the test molecule or molecules are incubated with the host cell in conjunction with one or more agonists held at a fixed concentration. An indication and relative strength of the antagonistic properties of the molecule(s) can be provided by comparing the level of gene expression in the indicator cell in the presence of the agonist, in
25 the absence of test molecules vs in the presence thereof. Of course, the antagonistic effect of a molecule can also be determined in the absence of agonist, simply by comparing the level of expression of the reporter gene product in the presence and absence of the test molecule(s).

It shall be understood that the "*in vivo*" experimental
30 model can also be used to carry out an "*in vitro*" assay. For example, cellular

extracts from the indicator cells can be prepared and used in an "*in vitro*" test. A non-limiting example thereof include binding assays.

As used herein the recitation "indicator cells" refers to cells that express a given genotype of AR according to the present invention. As alluded to above, such indicator cells can be used in the screening assays of the present invention. In certain embodiments, the indicator cells have been engineered so as to express a chosen derivative, fragment, homolog, or mutant of a genotype of the present invention. The cells can be yeast cells or higher eukaryotic cells such as mammalian cells. In one particular embodiment, the indicator cell would be a yeast cell harboring vectors enabling the use of the two hybrid system technology, as well known in the art (Ausubel et al., 1994, *supra*) and can be used to test a compound or a library thereof. In another embodiment, the *cis-trans* assay as described in USP 4,981,784, can be adapted and used in accordance with the present invention. Such an indicator cell could be used to rapidly screen at high-throughput a vast array of test molecules. In a particular embodiment, the reporter gene is luciferase or β -Gal.

In some embodiments, it might be beneficial to express a fusion protein. The design of constructs therefor and the expression and production of fusion proteins and are well known in the art (Sambrook et al., 1989, *supra*; and Ausubel et al., 1994, *supra*).

Non limiting examples of such fusion proteins include a hemagglutinin fusions and Gluthione-S-transferase (GST) fusions and Maltose binding protein (MBP) fusions. In certain embodiments, it might be beneficial to introduce a protease cleavage site between the two polypeptide sequences which have been fused. Such protease cleavage sites between two heterologously fused polypeptides are well known in the art.

In certain embodiments, it might also be beneficial to fuse the protein of the present invention to signal peptide sequences enabling a secretion of the fusion protein from the host cell. Signal peptides from diverse organisms are well known in the art. Bacterial OmpA and yeast Suc2 are two

non limiting examples of proteins containing signal sequences. In certain embodiments, it might also be beneficial to introduce a linker (commonly known) between the interaction domain and the heterologous polypeptide portion. Such fusion protein find utility in the assays of the present invention as well as for purification purposes, detection purposes and the like.

For certainty, the sequences and polypeptides useful to practice the invention include without being limited thereto mutants, homologs, subtypes, alleles and the like. It shall be understood that generally, the sequences of the present invention should encode a functional (albeit defective) AR. It will be clear to the person of ordinary skill that whether the AR sequence of the present invention, variant, derivative, or fragment thereof retains its function, can be determined by using the teachings and assays of the present invention and the general teachings of the art.

It should be understood that the AR protein of the present invention can be modified, for example by *in vitro* mutagenesis, to dissect the structure-function relationship thereof and permit a better design and identification of modulating compounds. However, some derivative or analogs having lost their biological function may still find utility, for example for raising antibodies. These antibodies could be used for detection or purification purposes. In addition, these antibodies could also act as competitive or non-competitive inhibitor and be found to be modulators of the activity of the AR protein of the present invention.

A host cell or indicator cell has been "transfected" by exogenous or heterologous DNA (e.g. a DNA construct) when such DNA has been introduced inside the cell. The transfecting DNA may or may not be integrated (covalently linked) into chromosomal DNA making up the genome of the cell. In prokaryotes, yeast, and mammalian cells for example, the transfecting DNA may be maintained on a episomal element such as a plasmid. With respect to eukaryotic cells, a stably transfected cell is one in which the transfecting DNA has become integrated into a chromosome so that it is

inherited by daughter cells through chromosome replication. This stability is demonstrated by the ability of the eukaryotic cell to establish cell lines or clones comprised of a population of daughter cells containing the transfecting DNA. Transfection methods are well known in the art (Sambrook et al., 1989, *supra*;
5 Ausubel et al., 1994 *supra*). The use of a mammalian cell as indicator can provide the advantage of furnishing an intermediate factor, which permits for example the interaction of two polypeptides which are tested, that might not be present in lower eukaryotes or prokaryotes. It will be understood that extracts from mammalian cells for example could be used in certain embodiments, to
10 compensate for the lack of certain factors.

In general, techniques for preparing antibodies (including monoclonal antibodies and hybridomas) and for detecting antigens using antibodies are well known in the art (Campbell, 1984, In "Monoclonal Antibody Technology: Laboratory Techniques in Biochemistry and Molecular Biology",
15 Elsevier Science Publisher, Amsterdam, The Netherlands) and in Harlow et al., 1988 (in: Antibody-A Laboratory Manual, CSH Laboratories). The present invention also provides polyclonal, monoclonal antibodies, or humanized versions thereof, chimeric antibodies and the like which inhibit or neutralize their respective interaction domains and/or are specific thereto.

20 From the specification and appended claims, the term therapeutic agent should be taken in a broad sense so as to also include a combination of at least two such therapeutic agents. Further, the DNA segments or proteins according to the present invention could be introduced into individuals in a number of ways. For example, cells can be isolated from the
25 afflicted individual, transformed with a DNA construct according to the invention and reintroduced to the afflicted individual in a number of ways. Alternatively, the DNA construct can be administered directly to the afflicted individual. The DNA construct can also be delivered through a vehicle such as a liposome, which can be designed to be targeted to a specific cell type, and engineered to be
30 administered through different routes. For example, an androgen receptor

gene having the genotype associated with low risk of breast cancer could be introduced in cells or in an individual displaying the AR polymorphism associated with high risk of breast cancer.

5 For administration to humans, the prescribing medical professional will ultimately determine the appropriate form and dosage for a given patient, and this can be expected to vary according to the chosen therapeutic regimen (i.e. DNA construct, protein, cells), the response and condition of the patient as well as the severity of the disease.

10 Composition within the scope of the present invention should contain the active agent (i.e. molecule, hormone) in an amount effective to achieve the desired therapeutic effect while avoiding adverse side effects. Typically, the nucleic acids in accordance with the present invention can be administered to mammals (i.e. humans) in doses ranging from 0.005 to 1 mg per kg of body weight per day of the mammal which is treated. Pharmaceutically
15 acceptable preparations and salts of the active agent are within the scope of the present invention and are well known in the art (Remington's Pharmaceutical Science, 16th Ed., Mack Ed.). For the administration of polypeptides, antagonists, agonists and the like, the amount administered should be chosen so as to avoid adverse side effects. The dosage will be adapted by the clinician
20 in accordance with conventional factors such as the extent of the disease and different parameters from the patient. Typically, 0.001 to 50 mg/kg/day will be administered to the mammal.

The present invention relates to a kit for assessing a predisposition to breast cancer comprising a determination of the genotype at
25 the AR locus (or a locus in linkage disequilibrium therewith) using a nucleic acid fragment, a protein or a ligand, or a restriction enzyme in accordance with the present invention. For example, a compartmentalized kit in accordance with the present invention includes any kit in which reagents are contained in separate containers. Such containers include small glass containers, plastic containers
30 or strips of plastic or paper. Such containers allow the efficient transfer of

reagents from one compartment to another compartment such that the samples and reagents are not cross-contaminated and the agents or solutions of each container can be added in a quantitative fashion from one compartment to another. Such containers will include in one particular embodiment a container which will accept the test sample (DNA protein or cells), a container which contains the primers used in the assay, containers which contain enzymes, containers which contain wash reagents, and containers which contain the reagents used to detect the extension products.

It will be readily recognized by the person of ordinary skill, that the nucleic acid sequences, probes, primers, antibodies and the like of the present invention enabling a detection of the CAG repeat polymorphism of the AR gene of the present invention can be incorporated into anyone of numerous established kit formats which are well known in the art.

Other objects, advantages and features of the present invention will become more apparent upon reading of the following non-restrictive description of preferred embodiments which is exemplary and should not be interpreted as limiting the scope of the present invention.

DESCRIPTION OF THE PREFERRED EMBODIMENT

In accordance with one embodiment of the invention, there is provided a specific model for use in prediction of breast cancer susceptibility and prognosis. The model comprises an androgen receptor gene polymorphism that allows to identify a subset of patients (i.e. women) that are at significantly increased risk of breast cancer as compared to those bearing other variants of this gene.

In accordance with a preferred embodiment of the present invention, a single gene, the androgen receptor gene, has been identified. The polymorphism of this gene is associated with a significant proportion of breast cancer cases in the general population (up to 60% of all cases). Polymorphism of this gene is for example the CAG repeat located in the first exon.

It was thus discovered in accordance with a preferred embodiment of the present invention that testing for this polymorphism in the androgen receptor (AR) gene allows to distinguish between women at lower risk of breast cancer and those at higher risk of the disease.

5 The present invention will be more readily understood by referring to the following example which is given to illustrate the invention rather than to limit its scope.

EXAMPLE 1

10 Polymorphism of the CAG repeat of the androgen receptor as a marker for breast cancer susceptibility

In a case control study comparing 262 consecutive cases of breast cancer in women and 465 control women matched for age, polymorphism at the AR gene, namely, the CAG repeat coding for a polyglutamine tract in the
15 5' part of the AR gene located on chromosome X, was studied. Because of the large number of alleles identified (15 different alleles), these alleles were grouped arbitrarily in categories by size to simplify the analysis and increase the number of individuals in each category. Table 1 presents the frequency of cases
20 and controls in categories of genotypes with the corresponding odds ratio for breast cancer and the computed 95% confidence intervals. The AR gene alleles were called arbitrarily A to E according to their size in CAG repeats, the shortest alleles being A and the longest being called E. The shortest AR gene alleles (corresponding to the polyglutamine stretch) or combinations of short alleles
25 (AA,AB,BB) are the genotypes that show the smallest breast cancer risk. This shows that women with a certain combination of AR gene polymorphisms on their two X chromosomes have a significantly increased risk of developing breast cancer as compared to the category with the smallest risk. In fact, in this cohort 32% of all cases of breast cancer were attributable to variation in the AR
30 gene. This is three to six times the number of breast cancer cases attributable

to the BRCA1 and BRCA2 genes. Indeed, in the cohort studied the 25% of women with the AR genotypes associated with the smallest risk of breast cancer comprised only 19% of all breast cancer cases while the 75% of women having the AR genotypes associated with the highest risks of breast cancer had 81% of all breast cancer cases. In other words, as compared with the general population, for which the risk of breast cancer is of 1:9 women, women with certain AR genotypes had a risk of 1:12 (much lower; i.e. protecting effect) while the other group had a risk of 1:8 (larger). Thus, this novel genetic marker of breast cancer allows to identify a subgroup of women with a risk of breast cancer close to two times larger than the other subgroup.

Table 1

Distribution of cases and controls among
females with various AR genotypes

5	AR genotype	Cases	Controls	Totals
	A* + BB	49	134	183
	BC to EE	213	331	544
	Totals	262	465	727

10 Odds Ratio (OR) for breast cancer in BC to EE genotypes vs A* and BB = 1.76 (95%
confidence interval CI 1.22 to 2.55)
Chi-square = 9.5 p=0.002
Breast cancer risk attributable to AR gene variation = 32% (57 cases / 213 in the others
category)

15

As will be clear to the skilled artisan, the different alleles AR
alleles can be grouped differently according to size, and the invention should
therefore not be limited to particular groupings. As will be seen in Table 2,
groupings of the alleles in three categories instead of 5, still enable a
20 demonstration of the significant association of the AR CAG-repeat polymorphism
with breast cancer.

In Table 2, the 15 different alleles were grouped in three
different categories (X, Y, and Z) instead of five, in which the shortest alleles are
in the X category, and the longest alleles are in the Z category. The six possible
25 genotypes were thus designated as "XX", "XY", "XZ", "YY", "YZ, and "ZZ"
genotypes. It is apparent from Table 2 that (CAG)_n genotypes were associated
with the disease as the genotypes with mid to large numbers of (CAG) repeats
were at significantly higher risk of developing the disease as compared to
genotypes with shorter (CAG)_n tracts (Table 2). Table 2 shows that women with
30 either the YY, YZ or ZZ genotypes had a 2.2-fold increased risk of breast cancer
compared to women with the XX or XY genotype, i.e. that women with these
later genotypes had only a 1:20 lifetime risk for the disease as compared to a
1:9 risk for those with the larger genotypes.

Table 2

Association of androgen receptor
polymorphism with breast cancer

	(CAG)n genotype		
	XX or XY genotype	XZ genotype	YY, YZ or ZZ genotype
Cases	10 (4%)*	28 (11%)*	212 (85%)*
Controls	37 (8%)*	61 (13%)*	355 (78%)*
Odds Ratio (OR)	1.0	1.7	2.2
95% CI for OR (min-max)		0.7 - 3.9	1.1 - 4.5
Lifetime risk of breast cancer	1:20	1:12	1:9

* value in parenthesis represents percentage of total cases or controls
CI Confidence interval expressed with the highest and lowest values.

No significant interaction was observed between AR genotypes and the body mass index (BMI), smoking habits, menopausal status or family history of breast cancer. However, a striking combined influence of the AR genotype with a positive history of breast benign disease (BBD) on the risk of breast cancer was observed (Table 3). Women with a positive history of BBD and AR genotypes combining the large AR alleles (Y or Z) had a relative risk of 3.5 as compared to women with no such history and AR genotypes comprised of smaller alleles. When compared to carriers of XX, XY AR genotypes only (who have the lowest risk of breast cancer) with no history of benign disease, women with the AR-ZZ genotype had an odds ratio of 7.1 for breast cancer (95% CI 2.3 to 22). Interestingly the AR genotype was not associated with a significant risk of breast cancer in women with no history of breast benign disease. The present invention thus also provides as an additional "marker" to

strengthen the prognosis/diagnosis/treatment methods and reagents according to the present invention, a positive history of BBD.

Table 3

Association of breast cancer risk with
AR polymorphism and breast benign disease

		AR genotype	
		XX, XY, XZ	YY, YZ, ZZ
10	Negative history of benign breast disease	Cases	25 (10%)*
		Controls	131 (52%)*
		Odds Ratio	1.33
		95% CI for OR (min - max)	0.8 - 2.2
		Lifetime risk of breast cancer	1:12
15	Positive history of benign breast disease	Cases	13 (5%)*
		Controls	81 (32%)*
		Odds Ratio	3.5
		95% CI for OR (min - max)	2.0 - 6.2
		Lifetime risk of breast cancer	1:4
20	Positive history of benign breast disease	Cases	25 (6%)*
		Controls	67 (15%)*
		Odds Ratio	1.5
		95% CI for OR (min - max)	0.7 - 3.4
		Lifetime risk of breast cancer	1:11

* value in parenthesis represents percentage of total cases or controls
CI Confidence interval expressed with the highest and lowest values.

Up to now, no marker displaying such a large odds ratios had been reported for breast cancer. Furthermore, this genetic marker and polymorphisms in the AR gene play a very significant role in breast cancer susceptibility in women, as evidenced by the very significant association demonstrated herein. The present invention also points to alternative therapies for breast cancer aiming at restoring the efficacy of the AR in women with a reduced function of their AR genes due to the variant genotypes that they carry. The described assays of the present invention could enables the identification of such therapies.

Although the present invention has been described hereinabove by way of preferred embodiments thereof, it can be modified, without departing from the spirit and nature of the subject invention as defined in the appended claims.

SEQUENCE LISTING

SEQ ID NO:1 5'-TCCAGAATCTGTTCCAGAGCGTGC-3

SEQ ID NO:2 5'-GCTGTGAAGGTTGCTGTTCTCAT-3'

WHAT IS CLAIMED IS:

1. A method of determining an individual's predisposition to breast cancer, development of breast cancer and/or responsiveness to therapy for breast cancer, said method comprising the step of determining the androgen receptor genotype of the individual, thereby determining an individual's predisposition to breast cancer, development of breast cancer and/or responsiveness to therapy for breast cancer.
2. The method of claim 1, wherein the androgen receptor genotype is determined using a nucleic acid variant in linkage disequilibrium with a CAG repeat in an androgen receptor gene.
3. The method of claim 2, wherein the androgen receptor genotype is determined by determining the number of CAG repeat within the androgen receptor gene
4. The method of claim 3, which further comprises a step of amplifying a segment of the androgen receptor using polymerase chain reaction.
5. The method of claim 4, wherein a pair of primers derived from a nucleic acid sequence of the androgen receptor gene or flanking said gene is used in the polymerase chain reaction.
6. The method of claim 5, wherein the segment of the androgen receptor gene is amplified using a pair of primers as follows:

5'-TCCAGAATCT GTTCCAGAGC GTGC-3' SEQ ID NO:1; and
5'-GCTGTGAAGG TTGCTGTTCC TCAT-3' SEQ ID NO:2.

7. An assay for screening and selecting an agent which
5 modulates breast cancer predisposition comprising:

a) a recombinant androgen receptor gene or functional
fragment thereof, which comprises the CAG repeat thereof, or a marker in
linkage disequilibrium therewith; and

b) assaying a function of said androgen receptor;

10 wherein an allele which modulates said function of an androgen receptor can be
selected,

and wherein a modulation of a function of said androgen receptor is associated
with a modulation of said breast cancer predisposition, whereby short CAG
repeat of said AR positively modulates AR receptor function, thereby leading to
15 breast cancer protection.

8. An assay for screening and selecting an agent which
modulates breast cancer predisposition comprising:

a) an expression vector comprising a promoter operably
20 linked to a reporter gene, said promoter comprising an androgen response
element, said response element affecting the activity of said promoter upon
binding thereto of androgen or analog thereof;

b) a cell expressing a chosen allele of an androgen receptor
and harboring said vector of a);

25 c) submitting said cell to at least one agent; and

d) assaying a level of said reporter gene;

whereby an agent can be selected when the level of said reporter gene is
significantly modulated by the presence of said agent.

9. A method of using specific alleles of the androgen receptor gene, or a variant, equivalent, or mutation thereof which shows linkage disequilibrium therewith, to set-up a screening assay for agents destined to modulate androgen receptor function for the purpose of identifying agents involved in breast cancer.
- 5

PCT

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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(21) International Application Number: PCT/CA99/00852 (22) International Filing Date: 15 September 1999 (15.09.99) (30) Priority Data: 60/100,311 15 September 1998 (15.09.98) US (71) Applicant (for all designated States except US): SIGNALGENE INC. [CA/CA]; 8475 Christophe Colomb Avenue, Montreal, Quebec H2M 2N9 (CA). (72) Inventor; and (75) Inventor/Applicant (for US only): ROUSSEAU, François [CA/CA]; 2540 Lalonde, Sainte-Foy, Quebec G1W 1M7 (CA). (74) Agents: DUBUC, Jean, H. et al.; Goudreau Gage Dubuc & Martineau Walker, The Stock Exchange Tower, Suite 3400, 800 Place Victoria, P.O. Box 242, Montreal, Quebec H4Z 1E9 (CA).		(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i> (88) Date of publication of the international search report: 25 May 2000 (25.05.00)
(54) Title: MARKER AT THE ANDROGEN RECEPTOR GENE FOR DETERMINING BREAST CANCER SUSCEPTIBILITY		
(57) Abstract <p>The present invention relates to a method of determining an individual's predisposition to breast cancer, development of breast cancer, protection against breast cancer and/or responsiveness to therapy for breast cancer. The method comprises the step of determining the androgen receptor genotype at the CAG repeat locus of an individual, or at a locus in linkage disequilibrium with the CAG repeat locus, thereby determining an individual's predisposition to breast cancer, development of breast cancer, protection against breast cancer and/or responsiveness to therapy for breast cancer.</p>		

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INTERNATIONAL SEARCH REPORT

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A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C12Q1/68

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B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 C12Q C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	ELHAJI Y ET AL: "The polymorphic CAG repeat of the androgen receptor and female breast cancer" AMERICAN JOURNAL OF HUMAN GENETICS, vol. 61, no. 4 suppl, 28 October 1997 (1997-10-28) - 1 November 1997 (1997-11-01), page A64 XP000884969 = see abstract 346 -/-	1-5

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INTERNATIONAL SEARCH REPORT

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A	WO 98 05797 A (LAMPARSKI HENRY G ;CALYDON (US); SCHUUR ERIC R (US); YU DE CHAO (U) 12 February 1998 (1998-02-12) page 8, line 4 - line 13	7-9

INTERNATIONAL SEARCH REPORT

Information on patent family members

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Patent document cited in search report		Publication date	Patent family member(s)	Publication date
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B. FIELDS SEARCHEDMinimum documentation searched (classification system followed by classification symbols)
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INTERNATIONAL SEARCH REPORT

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

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- (71) Applicant (*for all designated States except US*): UNIVERSITY OF ROCHESTER [US/US]; 601 Elmwood Avenue, Box 706, Rochester, NY 14642 (US).
- (72) Inventor; and
- (75) Inventor/Applicant (*for US only*): CHANG, Chawn-shang [US/US]; 19 Sandy Lane, Pittsford, NY 14534 (US).
- (74) Agents: SPRATT, Gwendolyn, D. et al.; Needle & Rosenberg, P.C., Suite 1200, The Candler Building, 127 Peachtree Street, N.E., Atlanta, GA 30303-1811 (US).
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WO 02/10452 A2

(54) Title: METHODS AND COMPOSITIONS FOR PREDICTING PROSTATE CANCER

(57) Abstract: Disclosed are compositions and methods for assessing risk in a subject for prostate cancer.

5 **METHODS AND COMPOSITIONS FOR PREDICTING** **PROSTATE CANCER**

 This application claims priority to United States Provisional Application No 60/221,074 filed on July 27, 2000, which application is herein incorporated by reference in its entirety.

10 **I. BACKGROUND OF THE INVENTION**

 The incidence of clinical prostate cancer differs substantially between ethnic groups, with African Americans having a 10- to 40-fold higher incidence than Asians (1-3AR). Such disparity in incidence of clinical prostate cancer cannot be explained entirely by population differences in screening. An earlier study shows
15 that after adjustment for screening, there is still a 3- to 4-fold difference in incidence rates between U.S. and Japanese men, whose rates are among the highest in Asians (4AR). Despite the dramatic racial variation in clinical prostate cancer incidence, the prevalence of latent carcinoma appears to be similar across populations (5AR), suggesting that there exists differences in factors (either genetic or environmental)
20 that promote the progression of microscopic tumors to clinically overt carcinoma.

 The growth, differentiation, and proliferation of prostatic cells are regulated by androgens (6AR). The biological effects of androgens are mediated through binding to the intracellular androgen receptor (AR), which in turn regulates the transcription of target genes with the assistance of transcriptional coactivators
25 (7AR).

II. SUMMARY OF THE INVENTION

 In accordance with the purposes of this invention, as embodied and broadly described herein, this invention, in one aspect, relates compositions and methods for assessing prostate cancer risk.

30 Additional advantages of the invention will be set forth in part in the description which follows, and in part will be obvious from the description, or may be learned by practice of the invention. The advantages of the invention will be realized and attained by means of the elements and combinations particularly pointed out in the appended claims. It is to be understood that both the foregoing
35 general description and the following detailed description are exemplary and

5 explanatory only and are not restrictive of the invention, as claimed.

III. BRIEF DESCRIPTION OF THE DRAWINGS

The accompanying drawings, which are incorporated in and constitute a part of this specification, illustrate several embodiments of the invention and together with the description, serve to explain the principles of the invention.

10 Figure 1. (AR) shows the percent distribution of the number of CAG repeats.

IV. DETAILED DESCRIPTION

The present invention may be understood more readily by reference to the following detailed description of preferred embodiments of the invention and the Examples included therein and to the Figures and their previous and following
15 description.

Before the present compounds, compositions, articles, devices, and/or methods are disclosed and described, it is to be understood that this invention is not limited to specific synthetic methods, specific recombinant biotechnology methods unless otherwise specified, or to particular reagents unless otherwise specified, as
20 such may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only and is not intended to be limiting.

A. Definitions

As used in the specification and the appended claims, the singular forms "a,"
25 "an" and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "a pharmaceutical carrier" includes mixtures of two or more such carriers, and the like.

Ranges may be expressed herein as from "about" one particular value, and/or to "about" another particular value. When such a range is expressed, another
30 embodiment includes from the one particular value and/or to the other particular value. Similarly, when values are expressed as approximations, by use of the antecedent "about," it will be understood that the particular value forms another embodiment. It will be further understood that the endpoints of each of the ranges

- 5 are significant both in relation to the other endpoint, and independently of the other endpoint.

In this specification and in the claims which follow, reference will be made to a number of terms which shall be defined to have the following meanings:

- 10 “Optional” or “optionally” means that the subsequently described event or circumstance may or may not occur, and that the description includes instances where said event or circumstance occurs and instances where it does not.

- 15 “Primers” are a subset of probes which are capable of supporting some type of enzymatic manipulation and which can hybridize with a target nucleic acid such that the enzymatic manipulation can occur. A primer can be made from any combination of nucleotides or nucleotide derivatives or analogs available in the art which do not interfere with the enzymatic manipulation.

- 20 “Probes” are molecules capable of interacting with a target nucleic acid, typically in a sequence specific manner, for example through hybridization. The hybridization of nucleic acids is well understood in the art and discussed herein. Typically a probe can be made from any combination of nucleotides or nucleotide derivatives or analogs available in the art.

B. Compositions

- 25 Disclosed are the components to be used to prepare the disclosed compositions as well as the compositions themselves and to be used within the methods disclosed herein. These and other materials are disclosed herein, and it is understood that when combinations, subsets, interactions, groups, etc. of these materials are disclosed that while specific reference of each various individual and collective permutation of these compounds may not be explicitly disclosed, each is specifically contemplated and described herein. For example, if a particular AIB1 30 protein or gene is disclosed and discussed and a number of modifications that can be made to a number of molecules including the AIB1 protein or gene are discussed, specifically contemplated is each and every combination and permutation of AIB1 protein or gene and the modifications that are possible unless specifically indicated

5 to the contrary. Thus, if a class of molecules A, B, and C are disclosed as well as a class of molecules D, E, and F and an example of a combination molecule, A-D is disclosed, then even if it each is not individually recited each is individually and collectively contemplated meaning combinations, A-E, A-F, B-D, B-E, B-F, C-D, C-E, and C-F are considered disclosed. Likewise, any subset or combination of these
10 is also disclosed. Thus, for example, the sub-group of A-E, B-F, and C-E would be considered disclosed. This concept applies to all aspects of this application including, but not limited to, steps in methods of making and using the disclosed compositions. Thus, if there are a variety of additional steps that can be performed it is understood that each of these additional steps can be performed with any specific
15 embodiment or combination of embodiments of the disclosed methods.

It is understood that one way to define any known variants and derivatives or those that might arise, of the disclosed genes and proteins herein is through defining the variants and derivatives in terms of homology to specific known sequences. For example, SEQ ID NO:18 sets forth a particular sequence of an AIB1 gene and SEQ
20 ID NO:19 sets forth a particular sequence of the protein encoded by SEQ ID NO:18, an AIB1 protein. Specifically disclosed are variants of these and other genes and proteins herein disclosed which have at least, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99 percent homology to the stated sequence. Those of skill in the art readily understand how to
25 determine the homology of two proteins or nucleic acids, such as genes. For example, the homology can be calculated after aligning the two sequences so that the homology is at its highest level.

Another way of calculating homology can be performed by published algorithms. Optimal alignment of sequences for comparison may be conducted by
30 the local homology algorithm of Smith and Waterman Adv. Appl. Math. 2: 482 (1981), by the homology alignment algorithm of Needleman and Wunsch, J. Mol. Biol. 48: 443 (1970), by the search for similarity method of Pearson and Lipman, Proc. Natl. Acad. Sci. U.S.A. 85: 2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics

- 5 Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI, or by inspection.

The same types of homology can be obtained for nucleic acids by for example the algorithms disclosed in Zuker, M. *Science* 244:48-52, 1989, Jaeger et al. *Proc. Natl. Acad. Sci. USA* 86:7706-7710, 1989, Jaeger et al. *Methods Enzymol.*
10 183:281-306, 1989 which are herein incorporated by reference for at least material related to nucleic acid alignment.

The term hybridization typically means a sequence driven interaction between at least two nucleic acid molecules, such as a primer or a probe and a gene. Sequence driven interaction means an interaction that occurs between two
15 nucleotides or nucleotide analogs or nucleotide derivatives in a nucleotide specific manner. For example, G interacting with C or A interacting with T are sequence driven interactions. Typically sequence driven interactions occur on the Watson-Crick face or Hoogsteen face of the nucleotide. The hybridization of two nucleic acids is affected by a number of conditions and parameters known to those of skill in
20 the art. For example, the salt concentrations, pH, and temperature of the reaction all affect whether two nucleic acid molecules will hybridize.

Parameters for selective hybridization between two nucleic acid molecules are well known to those of skill in the art. For example, in some embodiments selective hybridization conditions can be defined as stringent hybridization
25 conditions. For example, stringency of hybridization is controlled by both temperature and salt concentration of either or both of the hybridization and washing steps. For example, the conditions of hybridization to achieve selective hybridization may involve hybridization in high ionic strength solution (6X SSC or 6X SSPE) at a temperature that is about 12-25°C below the T_m (the melting
30 temperature at which half of the molecules dissociate from their hybridization partners) followed by washing at a combination of temperature and salt concentration chosen so that the washing temperature is about 5°C to 20°C below the T_m . The temperature and salt conditions are readily determined empirically in preliminary experiments in which samples of reference DNA immobilized on filters

5 are hybridized to a labeled nucleic acid of interest and then washed under conditions of different stringencies. Hybridization temperatures are typically higher for DNA-RNA and RNA-RNA hybridizations. The conditions can be used as described above to achieve stringency, or as is known in the art. (Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., Cold Spring Harbor Laboratory, Cold
10 Spring Harbor, New York, 1989; Kunkel et al. Methods Enzymol. 1987:154:367, 1987 which is herein incorporated by reference for material at least related to hybridization of nucleic acids). A preferable stringent hybridization condition for a DNA:DNA hybridization can be at about 68°C (in aqueous solution) in 6X SSC or 6X SSPE followed by washing at 68°C. Stringency of hybridization and washing, if
15 desired, can be reduced accordingly as the degree of complementarity desired is decreased, and further, depending upon the G-C or A-T richness of any area wherein variability is searched for. Likewise, stringency of hybridization and washing, if desired, can be increased accordingly as homology desired is increased, and further, depending upon the G-C or A-T richness of any area wherein high homology is
20 desired, all as known in the art.

Another way to define selective hybridization is by looking at the amount (percentage) of one of the nucleic acids bound to the other nucleic acid. For example, in some embodiments selective hybridization conditions would be when at least, 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 71, 72, 73, 74, 75, 76, 77,
25 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100 percent of the limiting nucleic acid is bound to the non-limiting nucleic acid. Typically, the non-limiting primer is in for example, 10 or 100 or 1000 fold excess. This type of assay can be performed at under conditions where both the limiting and non-limiting primer are for example, 10 fold or 100 fold or 1000 fold below their k_d ,
30 or where only one of the nucleic acid molecules is 10 fold or 100 fold or 1000 fold or where one or both nucleic acid molecules are above their k_d .

Another way to define selective hybridization is by looking at the percentage of primer that gets enzymatically manipulated under conditions where hybridization is required to promote the desired enzymatic manipulation. For example, in some

5 embodiments selective hybridization conditions would be when at least, 5, 10, 15,
20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81,
82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100 percent of
the primer is enzymatically manipulated under conditions which promote the
enzymatic manipulation, for example if the enzymatic manipulation is DNA
10 extension, then selective hybridization conditions would be when at least 5, 10, 15,
20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81,
82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100 percent of
the primer molecules are extended. Preferred conditions also include those
suggested by the manufacturer or indicated in the art as being appropriate for the
15 enzyme performing the manipulation.

1. AIB1/SRC3

The AIB1 protein (Amplified In Breast cancer 1, also known as Steroid
Receptor Coactivator-3, or SRC-3), encoded by the *AIB1/SRC-3* gene located on
chromosome 20 (20q12), is an AR coactivator and a member of the steroid receptor
20 coactivator family, which interacts with members of the nuclear hormone receptor
family (14,15 AIB1). Like the AR protein, the AIB1/SRC-3 coactivator contains a
stretch of glutamine residues encoded by a variable-length track of CAG/CAA
repeats in the *AIB1/SRC-3* gene.

a) Sequences

25 There are a variety of sequences related to the AIB1 gene having the
following Genbank Accession Numbers: NT_011362, XM_030033, XM_030032,
XM_009483, XM_030031, XM_030034, AL353777, AL034418, AL021394,
AY008258, AF322224, NM_008679, NM_006534, AF012108, and AF044080,
these sequences and others are herein incorporated by reference in their entireties as
30 well as for individual subsequences contained therein.

One particular sequence set forth in SEQ ID NO:18 and having Genbank
accession number XM_030032 is used herein, as an example, to exemplify the
disclosed compositions and methods. It is understood that the description related to
this sequence is applicable to any sequence related to AIB1 unless specifically

5 indicated otherwise. Those of skill in the art understand how to resolve sequence discrepancies and differences and to adjust the compositions and methods relating to a particular sequence to other related sequences (i.e. sequences of AIB1). Primers and/or probes can be designed for any AIB1 sequence given the information disclosed herein and known in the art.

10 b) CAG/CAA region

The CAG/CAA region of the AIB1 gene is located in the coding region of the AIB1 gene. For example, the CAG/CAA region in SEQ ID NO:18 can be defined by the region from nucleotide 3930 to nucleotide 4016. In certain embodiments of the disclosed compositions and methods, this represents the
15 CAG/CAA region of the AIB1 gene. However, it is understood that various mutations, alterations, or other genetic variation including allelic variation can occur in certain individuals and those of skill in the art understand how to locate this region within any given AIB1 gene variant. Thus, for example, if a nucleic acid is amplified from SEQ ID NO:18 or any variant of the AIB1 gene, that contains only
20 only a fragment of the AIB1 gene, for example, a fragment of 1000 nucleotides, it is understood that the CAG/CAA region could be located within this molecule if it is included in the molecule, in whole or in part.

c) Primers and probes

Disclosed are compositions including primers and probes, which are capable
25 of interacting with the AIB1 gene as disclosed herein. In certain embodiments the primers are used to support DNA amplification reactions. Typically the primers will be capable of being extended in a sequence specific manner. Extension of a primer in a sequence specific manner includes any methods wherein the sequence and/or composition of the nucleic acid molecule to which the primer is hybridized or
30 otherwise associated directs or influences the composition or sequence of the product produced by the extension of the primer. Extension of the primer in a sequence specific manner therefore includes, but is not limited to, PCR, DNA sequencing, DNA extension, DNA polymerization, RNA transcription, or reverse transcription. Techniques and conditions that amplify the primer in a sequence

5 specific manner are preferred. In certain embodiments the primers are used for the DNA amplification reactions, such as PCR or direct sequencing. It is understood that in certain embodiments the primers can also be extended using non-enzymatic techniques, where for example, the nucleotides or oligonucleotides used to extend the primer are modified such that they will chemically react to extend the primer in a
10 sequence specific manner. Typically the disclosed primers hybridize with the AIB1 gene or region of the AIB1 gene or they hybridize with the complement of the AIB1 gene or complement of a region of the AIB1 gene.

Disclosed are primers that are capable of amplifying the CAG/CAA region of the AIB1 gene. In certain embodiments the primers amplify the CAG/CAA region
15 of the AIB1 gene from nucleotide 3930 to nucleotide 4016 of the sequence set forth in SEQ ID NO:18. In certain embodiments the primers are "outside" the CAG/CAA region of the AIB1 gene. By outside the region it is intended to indicate that no region of the primer is intended to interact directly with the CAG/CAA region. For example, a primer outside the CAG/CAA region of the sequence set forth in SEQ ID
20 NO:18 could hybridize with nucleotide 4017 to nucleotide 4037 of SEQ ID NO:18, but it would not be intended to hybridize with nucleotide 4016, and under the conditions designed for the enzymatic manipulation of the primer, it would not appreciably interact with nucleotide 4016. In other embodiments the primers are designed to interact with one or more nucleotides considered to be part of the
25 CAG/CAA region, which primers herein are referred to as "inside" primers. Thus, an inside primer can include a primer region that can under conditions appropriate for the desired enzymatic manipulation hybridize with one or more nucleotides considered within the CAG/CAA region and contain a region that hybridizes with nucleotides that are not considered part of the CAG/CAA region. Thus, in one
30 embodiment an inside primer would interact with the nucleotide at position 4016 of SEQ ID NO:18.

The size of the primers for interaction with the AIB1 gene in certain embodiments can be any size that supports the desired enzymatic manipulation of the primer, such as DNA amplification. A typical AIB1 primer would be at least 6,

5 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29,
30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51,
52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73,
74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95,
96, 97, 98, 99, 100, 125, 150, 175, 200, 225, 250, 275, 300, 325, 350, 375, 400, 425,
10 450, 475, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1250, 1500, 1750,
2000, 2250, 2500, 2750, 3000, 3500, or 4000 nucleotides long.

In other embodiments an AIB1 primer can be less than or equal to 6, 7, 8, 9,
10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31,
32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53,
15 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75,
76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97,
98, 99, 100, 125, 150, 175, 200, 225, 250, 275, 300, 325, 350, 375, 400, 425, 450,
475, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1250, 1500, 1750,
2000, 2250, 2500, 2750, 3000, 3500, or 4000 nucleotides long.

20 In certain embodiments the primers are designed such that they are outside
primers whose nearest point of interaction with the AIB1 gene is within 0, 1, 2, 3, 4,
5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28,
29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50,
51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72,
25 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94,
95, 96, 97, 98, 99, 100, 125, 150, 175, or 200 nucleotides of the outermost defining
nucleotide of the CAG/CAA region or complement of the CAG/CAA region.

In certain embodiments the primers are designed such that they are outside
primers whose nearest point of interaction with the AIB1 gene is at least 0, 1, 2, 3, 4,
30 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28,
29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50,
51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72,
73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94,
95, 96, 97, 98, 99, 100, 125, 150, 175, or 200 nucleotides away from the outermost

- 5 defining nucleotide of the CAG/CAA region or complement of the CAG/CAA region.

For example, with respect to the AIB1 gene set forth in SEQ ID NO:18, certain embodiments of the primers would be designed such that they are outside primers whose nearest point of interaction with the AIB1 gene occurs at position

10 4017, 4018, 4019, 4020, 4021, 4022, 4023, 4024, 4025, 4026, 4027, 4028, 4029, 4030, 4031, 4032, 4033, 4034, 4035, 4036, 4037, 4038, 4039, 4040, 4041, 4042, 4043, 4044, 4045, 4046, 4047, 4048, 4049, 4050, 4051, 4052, 4053, 4054, 4055, 4056, 4057, 4058, 4059, 4060, 4061, 4062, 4063, 4064, 4065, 4066, 4067, 4068, 4069, 4070, 4071, 4072, 4073, 4074, 4075, 4076, 4077, 4078, 4079, 4080, 4081,

15 4082, 4083, 4084, 4085, 4086, 4087, 4088, 4089, 4090, 4091, 4092, 4093, 4094, 4095, 4096, 4097, 4098, 4099, 4100, 4101, 4102, 4103, 4104, 4105, 4106, 4107, 4108, 4109, 4110, 4111, 4112, 4113, 4114, 4115, 4116, 4117, 4142, 4167, 4192, 4217 of SEQ ID NO:18.

The primers for the AIB1 gene typically will be used to produce an amplified

20 DNA product that contains the CAG/CAA region of the AIB1 gene. In general, typically the size of the product will be such that the size can be accurately determined to within 3, or 2 or 1 nucleotides.

In certain embodiments this product is at least 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49,

25 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 125, 150, 175, 200, 225, 250, 275, 300, 325, 350, 375, 400, 425, 450, 475, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1250, 1500, 1750, 2000, 2250, 2500, 2750, 3000, 3500, or 4000 nucleotides long.

30 In other embodiments the product is less than or equal to 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90,

5 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 125, 150, 175, 200, 225, 250, 275, 300, 325, 350, 375, 400, 425, 450, 475, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1250, 1500, 1750, 2000, 2250, 2500, 2750, 3000, 3500, or 4000 nucleotides long.

2. Androgen receptor

10 Androgen receptor (AR) plays a key role in intraprostatic androgenic action. Within the prostate gland, testosterone is converted into dihydrotestosterone (DHT), a more potent androgen. DHT then binds to the AR to form an intracellular DHT-AR complex, which in turn modulates prostatic target genes to induce proliferation.

The AR protein, consisting of 918 amino acids and encoded singly by the AR
15 gene located on the X chromosome (Xq11-12), has three major functional domains: a transactivating amino-terminal domain, a DNA binding domain, and a ligand (steroid) binding domain (8AR). The open reading frame of the AR gene is separated over eight exons and has a length of 2,730 base pair (bp). The sequence encoding the large amino-terminal transactivating domain is found in the first exon;
20 the DNA binding domain is encoded by exons 2 and 3; and the information for the ligand binding domain is distributed over exons 4 to 8 (8AR).

The first exon of the AR gene contains two polymorphic trinucleotide repeat segments that encode polyglutamine and polyglycine tracts localized in the N-terminal transactivation domain of the AR protein. The polyglutamine tract is
25 encoded by a CAG trinucleotide repeat, and the polyglycine stretch by a GGN repeat. The number of CAG repeats ranges from about 8 to 35 repeats in normal individuals. Longer CAG repeat lengths appear to result in reduced AR transcriptional activity both *in vivo* and *in vitro* (9,10AR). Otherwise healthy men whose androgen receptor has a CAG repeat length at the long end of the normal
30 range (>28) have an increased incidence of impaired spermatogenesis and infertility (11AR), conditions that are extremely androgen-dependent (12AR). Expansion of the CAG repeat length to over 40 repeats is related to a rare neuromuscular disorder, spinal and bulbar muscular atrophy (Kennedy syndrome), which is also associated with androgen insensitivity, decreased virilization, testicular atrophy, reduced sperm

5 production, and infertility (13-15AR). Together, these clinical data suggest that a longer CAG repeat length decreases the functional competence of AR.

The length of the polyglycine (GGN) tract varies from about 10 to 30 repeats. The functional consequences of variation in the GGN tract are less clear. Deletion of the polyglycine tract reduces AR transcriptional activity by about 30% in transient
10 transfection assays (16AR), although there is no significant correlation between polyglycine tract length and infertility (11AR).

a) Sequences

There are a variety of sequences related to the androgen receptor gene having the following Genbank Accession Numbers: NM_000044 or L49399 are two
15 examples. These sequences and others available on Genbank are herein incorporated by reference in their entirety as well as for individual subsequences contained therein.

One particular sequence set forth in SEQ ID NO:20 and having Genbank accession number NM_000044 is used herein, as an example, to exemplify the
20 disclosed compositions and methods. It is understood that the description related to this sequence is applicable to any sequence related to androgen receptor unless specifically indicated otherwise. Those of skill in the art understand how to resolve sequence discrepancies and differences and to adjust the compositions and methods relating to a particular sequence to other related sequences. Primers and/or probes
25 can be designed for any androgen receptor sequence given the information disclosed herein and known in the art.

b) CAG/CAA region

The CAG/CAA region in SEQ ID NO:20 can be defined by the region from nucleotide 1286 to nucleotide 1348. In certain embodiments of the disclosed
30 compositions and methods, this represents the CAG/CAA region of the androgen receptor gene. However, it is understood that various mutations, alterations, or other genetic variation including allelic variation can occur in certain individuals and those of skill in the art understand how to locate this region within any given

5 androgen receptor gene variant. Thus, for example, if a nucleic acid is amplified from SEQ ID NO:20 or any variant of the androgen receptor gene, that contains only 1000 nucleotides, it is understood that the CAG/CAA region could be located within this molecule if it is included in the molecule, in whole or in part.

c) GGN region

10 The GGN region in SEQ ID NO:20 can be defined by the region from nucleotide 2459 to nucleotide 2530. In certain embodiments of the disclosed compositions and methods, this represents the GGN region of the androgen receptor gene. However, it is understood that various mutations, alterations, or other genetic variation including allelic variation can occur in certain individuals and those of skill
15 in the art understand how to locate this region within any given androgen receptor gene variant. Thus, for example, if a nucleic acid is amplified from SEQ ID NO:20 or any variant of the androgen receptor gene, that contains only 1000 nucleotides, it is understood that the GGN region could be located within this molecule if it is included in the molecule, in whole or in part.

d) Primers and probes

20 Disclosed are compositions including primers and probes, which are capable of interacting with the androgen receptor gene as disclosed herein. In certain embodiments the primers are used to support DNA amplification reactions. Typically the primers will be capable of being extended in a sequence specific
25 manner. Extension of a primer in a sequence specific manner includes any methods wherein the sequence and/or composition of the nucleic acid molecule to which the primer is hybridized or otherwise associated directs or influences the composition or sequence of the product produced by the extension of the primer. Extension of the primer in a sequence specific manner therefore includes, but is not limited to, PCR,
30 DNA sequencing, DNA extension, DNA polymerization, RNA transcription, or reverse transcription. Techniques and conditions that amplify the primer in a sequence specific manner are preferred. In certain embodiments the primers are used for the DNA amplification reactions, such as PCR or direct sequencing. It is understood that in certain embodiments the primers can also be extended using non-

5 enzymatic techniques, where for example, the nucleotides or oligonucleotides used to extend the primer are modified such that they will chemically react to extend the primer in a sequence specific manner. Typically the disclosed primers hybridize with the androgen receptor gene or region of the androgen receptor gene or they hybridize with the complement of the androgen receptor gene or complement of a
10 region of the androgen receptor gene.

Disclosed are primers that are capable of amplifying the CAG/CAA and/or GGN region of the androgen receptor gene. In certain embodiments the primers amplify the CAG/CAA region of the androgen receptor gene from nucleotide 1286 to nucleotide 1348 of the sequence set forth in SEQ ID NO:20. In certain
15 embodiments the primers are "outside" the CAG/CAA region of the androgen receptor gene. By outside the region it is intended to indicate that no region of the primer is intended to interact directly with the CAG/CAA region. For example, a primer outside the CAG/CAA region of the sequence set forth in SEQ ID NO:20 could hybridize with nucleotide 1349 to nucleotide 1369 of SEQ ID NO:20, but it
20 would not be intended to hybridize with nucleotide 1348, and under the conditions designed for the enzymatic manipulation of the primer, it would not appreciably interact with nucleotide 1348. In other embodiments the primers are designed to interact with one or more nucleotides considered to be part of the CAG/CAA region, which primers herein are referred to as "inside" primers. Thus, an inside primer can
25 include a primer region that can under conditions appropriate for the desired enzymatic manipulation hybridize with one or more nucleotides considered within the CAG/CAA region and contain a region that hybridizes with nucleotides that are not considered part of the CAG/CAA region. Thus, in one embodiment an inside primer would interact with the nucleotide at position 1348 of SEQ ID NO:20.

30 In certain embodiments the primers amplify the GGN region of the androgen receptor gene from nucleotide 2459 to nucleotide 2530 of the sequence set forth in SEQ ID NO:20. In certain embodiments the primers are "outside" the GGN region of the androgen receptor gene. By outside the region it is intended to indicate that no region of the primer is intended to interact directly with the GGN region. For

5 example, a primer outside the GGN region of the sequence set forth in SEQ ID NO:20 could hybridize with nucleotide 2531 to nucleotide 2551 of SEQ ID NO:20, but it would not be intended to hybridize with nucleotide 2530, and under the conditions designed for the enzymatic manipulation of the primer, it would not appreciably interact with nucleotide 2530. In other embodiments the primers are
10 designed to interact with one or more nucleotides considered to be part of the GGN region, which primers herein are referred to as "inside" primers. Thus, an inside primer can include a primer region that can under conditions appropriate for the desired enzymatic manipulation hybridize with one or more nucleotides considered within the GGN region and contain a region that hybridizes with nucleotides that are
15 not considered part of the GGN region. Thus, in one embodiment an inside primer would interact with the nucleotide at position 2530 of SEQ ID NO:20.

The size of the primers for interaction with the androgen receptor gene in certain embodiments can be any size that supports the desired enzymatic manipulation of the primer, such as DNA amplification. A typical androgen
20 receptor primer would be at least 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 125, 150, 175, 200, 225, 250,
25 275, 300, 325, 350, 375, 400, 425, 450, 475, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1250, 1500, 1750, 2000, 2250, 2500, 2750, 3000, 3500, or 4000 nucleotides long.

In other embodiments an androgen receptor primer can be less than or equal to 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28,
30 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 125, 150, 175, 200, 225, 250, 275, 300, 325, 350, 375, 400, 425, 450, 475, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1250, 1500,

5 1750, 2000, 2250, 2500, 2750, 3000, 3500, or 4000 nucleotides long.

In certain embodiments the primers are designed such that they are outside primers whose nearest point of interaction with the androgen receptor gene is within 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 10 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 125, 150, 175, or 200 nucleotides of the outermost defining nucleotide of the CAG/CAA and/or GGN region or complement of the CAG/CAA and/or GGN region.

15 In certain embodiments the primers are designed such that they are outside primers whose nearest point of interaction with the androgen receptor gene is at least 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 20 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 125, 150, 175, or 200 nucleotides away from the outermost defining nucleotide of the CAG/CAA and/or GGN region or complement of the CAG/CAA and/or GGN region.

For example, with respect to the androgen receptor gene set forth in SEQ ID 25 NO:20, certain embodiments of the primers would be designed such that they are outside primers whose nearest point of interaction with the androgen receptor gene occurs at position 1349, 1350, 1351, 1352, 1353, 1354, 1355, 1356, 1357, 1358, 1359, 1360, 1361, 1362, 1363, 1364, 1365, 1366, 1367, 1368, 1369, 1370, 1371, 1372, 1373, 1374, 1375, 1376, 1377, 1378, 1379, 1380, 1381, 1382, 1383, 1384, 30 1385, 1386, 1387, 1388, 1389, 1390, 1391, 1392, 1393, 1394, 1395, 1396, 1397, 1398, 1399, 1400, 1401, 1402, 1403, 1404, 1405, 1406, 1407, 1408, 1409, 1410, 1411, 1412, 1413, 1414, 1415, 1416, 1417, 1418, 1419, 1420, 1421, 1422, 1423, 1424, 1425, 1426, 1427, 1428, 1429, 1430, 1431, 1432, 1433, 1434, 1435, 1436, 1437, 1438, 1439, 1440, 1441, 1442, 1443, 1444, 1445, 1446, 1447, 1448, 1449,

5 1474, 1499, 1524, or 1549 of SEQ ID NO:20.

The primers for the androgen receptor gene typically will be used to produce an amplified DNA product that contains the CAG/CAA and/or GGN region of the androgen receptor gene. In general, typically the size of the product will be such that the size can be accurately determined to within 3, or 2 or 1 nucleotides.

10 In certain embodiments this product is at least 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 125, 150, 175, 200, 225, 250, 275, 300, 325, 350, 375,
15 400, 425, 450, 475, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1250, 1500, 1750, 2000, 2250, 2500, 2750, 3000, 3500, or 4000 nucleotides long.

In other embodiments the product is less than or equal to 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90,
20 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 125, 150, 175, 200, 225, 250, 275, 300, 325, 350, 375, 400, 425, 450, 475, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1250, 1500, 1750, 2000, 2250, 2500, 2750, 3000, 3500, or 4000 nucleotides long.

25 3. Kits

Disclosed herein are kits that are drawn to reagents that can be used in practicing the methods disclosed herein. The kits can include any reagent or combination of reagent discussed herein or that would be understood to be required or beneficial in the practice of the disclosed methods. For example, the kits could
30 include primers to perform the amplification reactions discussed in certain embodiments of the methods, as well as the buffers and enzymes required to use the primers as intended. For example, disclosed is a kit for assessing a subject's risk for acquiring prostate cancer, comprising the oligonucleotides set forth in SEQ ID Nos:

5 1 and 2.

C. Methods of making the compositions

The compositions disclosed herein and the compositions necessary to perform the disclosed methods can be made using any method known to those of skill in the art for that particular reagent or compound unless otherwise specifically
10 noted. For example, the nucleic acids, such as, the oligonucleotides to be used as primers can be made using standard chemical synthesis methods or can be produced using enzymatic methods or any other known method. Such methods can range from standard enzymatic digestion followed by nucleotide fragment isolation (see for example, Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, 2nd
15 Edition (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1989) Chapters 5, 6) to purely synthetic methods, for example, by the cyanoethyl phosphoramidite method using a Milligen or Beckman System 1Plus DNA synthesizer (for example, Model 8700 automated synthesizer of Milligen-Bioscience, Burlington, MA or ABI Model 380B). Synthetic methods useful for making
20 oligonucleotides are also described by Ikuta *et al.*, *Ann. Rev. Biochem.* 53:323-356 (1984), (phosphotriester and phosphite-triester methods), and Narang *et al.*, *Methods Enzymol.*, 65:610-620 (1980), (phosphotriester method). Protein nucleic acid molecules can be made using known methods such as those described by Nielsen *et al.*, *Bioconjug. Chem.* 5:3-7 (1994).

25 **D. Methods of using the compositions**

It is understood that any variation or implementation of the compositions discussed herein is understood to be a particular embodiment which can be used and is described as being used in the disclosed methods. Disclosed are methods for assessing the risk that a subject may acquire prostate cancer. Methods are also
30 disclosed for assessing the clinical significance of the prostate cancer that a subject will get or already has. These methods involve assaying the length of regions of genes that have been shown herein to be related to the onset and severity of prostate cancer. The methods involve assaying regions of both the AIB1 gene and the AR gene either individually or collectively. There are particular regions of the AIB1

5 gene and the AR gene which have been shown to be related to the onset and severity of prostate cancer and these regions are found in all forms and variants of these genes. In one embodiment of the disclosed methods the length of the CAG/CAA repeat in either the AIB1 gene or AR gene of a subject, discussed above, is determined. This length can then be compared to known lengths, which have been
10 shown to be correlated with the onset and severity of prostate cancer. For example, in the AIB1 gene, the CAG/CAA region determined to be greater than, less than, or equal to 29 repeats. When the number of repeats present in the repeat region of the subject is less than or more than 29 at least one allele of the subject's AIB1 gene, this person is determined to have an increased risk of developing prostate cancer. It
15 is understood that since the AIB1 gene is present on chromosome 20, subjects will have two alleles of this gene. When a single allele has a length different than 29 the subject has an increased risk of acquiring prostate cancer. If, however, the subject has two alleles of the AIB1 gene that both have lengths in the CAG/CAA repeat region different than 29, the subject has a higher risk of acquiring prostate cancer
20 than if only one allele has a CAG/CAA length different than 29. Thus, if the alleles of the AIB1 gene are assayed individually, if the first allele assayed has a CAG/CAA repeat length different than 29, a certain amount of information as to the person's susceptibility will be gained, however, if the allele contains a CAG/CAA repeat equal to 29, the second allele must be looked at to get meaningful information as to
25 the person's likelihood of acquiring prostate cancer. Furthermore, more information will be gained by assaying the second allele even if the first allele has a CAG/CAA length less than or greater than 29 because the second allele may also have a CAG/CAA length different 29 and this provides more information about the subject's prostate cancer susceptibility. Thus, it is preferred that both alleles of the
30 AIB1 gene are assayed when performing the methods herein.

The CAG/CAA region of the subject's AR gene can also be assayed. However, in the AR length the length of the subject's CAG/CAA region in the AR gene is compared to 23 repeats. If there are more or less than 23 repeats the subject has an increased risk of prostate cancer. Furthermore, the risk of prostate cancer in a

5 subject is directly correlated with the length of the subject's CAG/CAA repeat. The greater the deviation of the number of CAG/CAA repeats in a subjects DNA from 23 repeats the greater the likelihood that the person will acquire prostate cancer.

1. Methods assaying the CAG/CAA repeat of the AIB1 gene

Methods are disclosed where only the length of CAG/CAA repeat in a
10 subject's AIB1 gene is assayed. Disclosed are methods for assessing the risk of developing prostate cancer in a human by analyzing the AIB1 gene of the subject.

Disclosed are methods for assessing the risk of prostate cancer in a human subject comprising determining the length of the contiguous CAG or CAA repeats in both AIB1 gene alleles of the subject and assessing whether the length of the CAG
15 or CAA repeats is less than, equal to, or greater than 29 repeats, a length less than or greater than 29 repeats in both alleles indicating an increased risk of prostate cancer in the subject.

Also disclosed are methods where determining the length of the repeats comprises amplifying a region of both AIB1 gene alleles comprising the contiguous
20 CAG or CAA repeat. Any type of amplification method can be employed to amplify the target regions. Preferred methods include PCR and direct sequencing of the target repeat regions.

When two repeat regions of two alleles are amplified two products will be produced. These products can then be assayed for their length, by for example, gel
25 chromatography, HPLC, or capillary gel electrophoresis. Thus, disclosed are methods that produce two PCR products and methods where the PCR products are analyzed by chromatography, including but not limited to gel electrophoresis.

In certain embodiments the sequence of the repeat region can be determined by, for example, direct sequencing of the repeat region. In some embodiments the
30 PCT product or other DNA amplification product, can itself be sequenced.

In certain embodiments of the disclosed methods, the repeat region is assayed or amplified by targeting primers or probes to the region or the areas of the target gene surrounding the region. These primers, for example, can be used to

5 amplify the region by for example, PCR. It is preferred that the method utilize primers as they are discussed herein. For example, the PCR product can be produced using a first AIB1 primer that selectively hybridizes with the complement of sequence 5' to the repeat region and a second AIB1 primer that selectively hybridizes with sequence 3' to the repeat region.

10 In certain embodiments, the PCR product can be produced using a first AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:3 and a second AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:4. In still other embodiments of the disclosed methods, the first AIB1 primer has the sequence set forth in SEQ ID NO:1 and the second AIB1 primer has the
15 sequence set forth in SEQ ID NO:2.

In certain embodiments determining how many CAG or CAA repeats there are comprises sequencing the CAG or CAA repeats.

2. Methods assaying the CAG/CAA repeat of the Androgen receptor gene

20 Just as with the AIB1 gene, there are repeat regions within the androgen receptor gene that can be assayed and used to assess a person's risk for acquiring prostate cancer. There are two repeat regions that can be assayed within the androgen receptor gene. The CAG/CAA repeat region within the AR gene can be assayed as discussed for the CAG/CAA repeat region of the AIB1 gene. There is a
25 key difference, however, and that is the length of the subject's CAG/CAA repeat region in the AR gene is compared to 23 repeats, not 29 repeats. The predictability of acquiring prostate cancer arises from a comparison to 23 CAG/CAA repeats. Furthermore, the CAG/CAA repeat within the AR gene is predictive based on single repeat changes. Thus, the likelihood of acquiring prostate cancer increases for each
30 difference in length from 23 repeats. Thus, for example, a subject having 15 repeats is more likely to acquire prostate cancer or have a clinically significant prostate cancer than a person having 16 repeats and a subject having 16 repeats is more likely to acquire prostate cancer or have a clinically significant prostate cancer than a person having 17 repeats and a subject having 17 repeats is more likely to acquire

- 5 prostate cancer or have a clinically significant prostate cancer than a person having 16 repeats and so forth. This relationship holds for all lengths of CAG/CAA repeats in the AR gene.

The AR gene also has a GGN repeat region whose length is related to the susceptibility of prostate cancer. The methods and reagents disclosed for the
10 CAG/CAA repeats of the AR gene apply to the GGN repeat within the AR gene and are all specifically contemplated herein.

3. Methods assaying the CAG/CAA repeat of the Androgen receptor gene and the AIB1 gene

- Methods are also disclosed where both the AIB1 gene and the AR gene are
15 assayed to arrive at a likelihood that a subject will acquire prostate cancer. It is understood that all of the conditions and permutations of performing the methods that involve assaying either AIB1 or AR alone can also be used and applied when both AIB1 and AR are assayed.

- Disclosed are methods for assessing the risk of developing prostate cancer in
20 a human by analyzing the AIB1 gene and the AR gene of the subject together.

- For example, disclosed are methods for assessing the risk of prostate cancer in a human subject comprising determining the length of the contiguous CAG or CAA repeats in the AIB1 gene alleles of the subject and assessing whether the length of the CAG or CAA repeats in each allele is less than, equal to, or greater
25 than 29 repeats, and determining the length of the contiguous CAG or CAA repeats in the androgen receptor gene of the subject and assessing whether the length of the CAG or CAA repeats is less than, equal to, or greater than 23 repeats, a length in at least one allele less than or greater than 29 repeats in the AIB1 gene and less than or greater than 23 repeats in the androgen receptor gene indicating an increased risk of
30 prostate cancer in the subject.

Also disclosed are methods wherein determining the length of the contiguous CAG or CAA repeats in the AIB1 gene alleles comprises amplifying a region of the AIB1 gene alleles comprising the CAG or CAA repeats and wherein determining the

- 5 length of the CAG or CAA repeats in the androgen receptor gene comprises
amplifying a region of the androgen receptor gene comprising the CAG or CAA
repeats.

Furthermore, methods are disclosed wherein the amplification of the regions
of the AIB1 gene and the androgen receptor gene is by PCR that produces a first and
10 a second AIB1 PCR product and an androgen receptor PCR product.

Disclosed are methods further comprising analyzing the PCR products by
chromatography and methods wherein the chromatography is gel electrophoresis.
The sequence of the PCR products can be determined.

The methods involving amplifying repeat regions in both AIB1 and the AR
15 genes can for example involve primers. It is understood that primers specific for
each repeat region would be used. For example, the AIB1 PCR product is produced
using a first AIB1 primer that selectively hybridizes with the sequence set forth in
SEQ ID NO:3 and a second AIB1 primer that selectively hybridizes with the
sequence set forth in SEQ ID NO:4. Additional methods are wherein the first AIB1
20 primer has the sequence set forth in SEQ ID NO:1 and the second AIB1 primer has
the sequence set forth in SEQ ID NO:2.

Or when assaying both genes the androgen receptor PCR product is produced
using a first androgen receptor CAG primer that selectively hybridizes with the
sequence set forth in SEQ ID NO:9 and a second androgen receptor CAG primer
25 that selectively hybridizes with the sequence set forth in SEQ ID NO:10. In some
embodiments the first androgen receptor CAG primer has the sequence set forth in
SEQ ID NO:7 and the second androgen receptor CAG primer has the sequence set
forth in SEQ ID NO:8.

Or when both the AIB1 and the AR genes are assayed, the AIB1 PCR
30 product is produced using a first AIB1 primer that selectively hybridizes with the
sequence set forth in SEQ ID NO:3 and a second AIB1 primer that selectively
hybridizes with the sequence set forth in SEQ ID NO:4. and wherein the androgen
receptor PCR product is produced using a first androgen receptor CAG primer that

- 5 selectively hybridizes with the sequence set forth in SEQ ID NO:9 and a second androgen receptor CAG primer that selectively hybridizes with the sequence set forth in SEQ ID NO:10.

In certain embodiments the first AIB1 primer has the sequence set forth in SEQ ID NO:1 and the second AIB1 primer has the sequence set forth in SEQ ID
10 NO:2 and wherein the first androgen receptor CAG primer has the sequence set forth in SEQ ID NO:7 and the second androgen receptor CAG primer has the sequence set forth in SEQ ID NO:8.

In some embodiments the methods are performed wherein more than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the
15 person and more than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

In other embodiments the methods are performed wherein more than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and less than 23 contiguous CAG or CAA repeats in the androgen receptor
20 gene of the person indicates an increased risk of prostate cancer in the subject.

In some embodiments the methods are performed wherein less than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and more than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

25 In other embodiments the methods are performed wherein less than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and less than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

In other embodiments the methods are performed wherein more or less than
30 29 contiguous CAG or CAA repeats in both alleles of the AIB1 gene of the person and more than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

5 In other embodiments the methods are performed wherein more or less than 29 contiguous CAG or CAA repeats in both alleles of the AIB1 gene of the person and less than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

10 In other embodiments the methods are performed wherein more than 29 contiguous CAG or CAA repeats in both alleles of the AIB1 gene of the person and more than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

15 In other embodiments the methods are performed wherein less than 29 contiguous CAG or CAA repeats in both alleles of the AIB1 gene of the person and less than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

20 Also disclosed are methods for assessing the risk of prostate cancer in a human subject comprising determining the length of the contiguous CAG or CAA repeats in the AIB1 gene alleles of the subject and assessing whether the length of the CAG or CAA repeats in each allele is less than, equal to, or greater than 29 repeats, and determining the length of the contiguous GGN repeats in the androgen receptor gene of the subject and assessing whether the length of the GGN repeats is less than, equal to, or greater than 23 repeats, a length in at least one allele less than or greater than 29 repeats in the AIB1 gene and less than or greater than 23 repeats in the androgen receptor gene indicating an increased risk of prostate cancer in the subject, wherein N is either T, G, or C.

30 Disclosed are methods wherein determining the length of the contiguous CAG or CAA repeats in the AIB1 gene alleles comprises amplifying a region of the AIB1 gene alleles comprising the CAG or CAA repeats and wherein determining the length of the contiguous GGN repeats in the androgen receptor gene comprises amplifying a region of the androgen receptor gene comprising the GGN repeats.

 Further methods are disclosed wherein the amplification of the regions of the AIB1 gene and the androgen receptor gene is by PCR that produces a first and a

- 5 second AIB1 PCR product and an androgen receptor PCR product.

In other methods the amplification further comprises analyzing the PCR products by chromatography.

The amplification products including the PCR products can be analyzed by gel electrophoresis.

- 10 In other embodiments the sequence of the PCR products is determined.

Disclosed are amplification methods wherein the AIB1 PCR product is produced using a first AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:3 and a second AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:4.

- 15 Also disclosed are amplification methods wherein the first AIB1 primer has the sequence set forth in SEQ ID NO:1 and the second AIB1 primer has the sequence set forth in SEQ ID NO:2.

- Disclosed are methods wherein the androgen receptor PCR product is produced using a first androgen receptor GGN primer that selectively hybridizes with the sequence set forth in SEQ ID NO:13 and a second androgen receptor GGN primer that selectively hybridizes with the sequence set forth in SEQ ID NO:14.
- 20

Also disclosed are methods wherein the first androgen receptor GGN primer has the sequence set forth in SEQ ID NO:11 and the second androgen receptor GGN primer has the sequence set forth in SEQ ID NO:12.

- 25 Further disclosed are methods wherein the AIB1 PCR product is produced using a first AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:3 and a second AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:4. and wherein the androgen receptor PCR product is produced using a first androgen receptor GGN primer that selectively hybridizes with the sequence set forth in SEQ ID NO:13 and a second androgen receptor GGN primer that selectively hybridizes with the sequence set forth in SEQ ID NO:14.
- 30

Also disclosed are methods wherein the first AIB1 primer has the sequence

- 5 set forth in SEQ ID NO:1 and the second AIB1 primer has the sequence set forth in SEQ ID NO:2 and wherein the first androgen receptor GGN primer has the sequence set forth in SEQ ID NO:11 and the second androgen receptor GGN primer has the sequence set forth in SEQ ID NO:12.

- 10 In some embodiments the methods are performed wherein more than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and more than 23 contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

- 15 In other embodiments the methods are performed wherein more than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and less than 23 contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

- 20 In some embodiments the methods are performed wherein less than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and more than 23 contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

In other embodiments the methods are performed wherein less than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and less than 23 contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

- 25 In some embodiments the methods are performed wherein more or less than 29 contiguous CAG or CAA repeats in both alleles of the AIB1 gene of the person and more than 23 contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

- 30 In other embodiments the methods are performed wherein more or less than 29 contiguous CAG or CAA repeats in both alleles of the AIB1 gene of the person and less than 23 contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

5 In other embodiments the methods are performed wherein more than 29 contiguous CAG or CAA repeats in both alleles of the AIB1 gene of the person and more than 23 contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

10 In other embodiments the methods are performed wherein less than 29 contiguous CAG or CAA repeats in both alleles of the AIB1 gene of the person and less than 23 contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

15 Also disclosed are methods for assessing the risk of prostate cancer in a human subject comprising determining the length of the contiguous CAG or CAA repeats in an AIB1 gene of the subject and assessing whether the length of the CAG repeats is less than, equal to, or greater than 29 repeats, a length less than or greater than 29 repeats indicating an increased risk of prostate cancer in the subject.

20 Methods for assessing the risk of prostate cancer in a human subject comprising determining the length of the contiguous CAG or CAA repeats in an AIB1 gene of the subject and assessing whether the length of the CAG or CAA repeats is less than, equal to, or greater than 29 repeats, and determining the length of the contiguous CAG or CAA repeats in the androgen receptor gene of the subject and assessing whether the length of the CAG or CAA repeats is less than, equal to, or greater than 23 repeats, a length less than or greater than 29 repeats in the AIB1
25 allele and less than or greater than 23 repeats in the androgen receptor gene indicating an increased risk of prostate cancer in the subject are also disclosed.

30 Disclosed are methods for assessing the risk of prostate cancer in a human subject comprising determining the length of the contiguous CAG or CAA repeats in an AIB1 gene of the subject and assessing whether the length of the CAG or CAA repeats is less than, equal to, or greater than 29 repeats, and determining the length of the contiguous GGN repeats in the androgen receptor gene of the subject and assessing whether the length of the GGN repeats is less than, equal to, or greater than 23 repeats, a length less than or greater than 29 repeats in the AIB1 allele and

- 5 less than or greater than 23 repeats in the androgen receptor gene indicating an increased risk of prostate cancer in the subject, wherein N is either T, G, or C.

Throughout this application, various publications are referenced. The disclosures of these publications in their entireties are hereby incorporated by reference into this application in order to more fully describe the state of the art to
10 which this invention pertains. The references disclosed are also individually and specifically incorporated by reference herein for the material contained in them that is discussed in the sentence in which the reference is relied upon.

It will be apparent to those skilled in the art that various modifications and variations can be made in the present invention without departing from the scope or
15 spirit of the invention. Other embodiments of the invention will be apparent to those skilled in the art from consideration of the specification and practice of the invention disclosed herein. It is intended that the specification and examples be considered as exemplary only, with a true scope and spirit of the invention being indicated by the following claims.

20 *E. Examples*

The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how the compounds, compositions, articles, devices and/or methods claimed herein are made and evaluated, and are intended to be purely exemplary of the invention and are not
25 intended to limit the scope of what the inventors regard as their invention. Efforts have been made to ensure accuracy with respect to numbers (e.g., amounts, temperature, etc.), but some errors and deviations should be accounted for. Unless indicated otherwise, parts are parts by weight, temperature is in °C or is at ambient temperature, and pressure is at or near atmospheric.

30 1. Example 1

Because AR coactivators enhance transactivation of AR, the relationship of a CAG/CAA repeat length polymorphism in the *AIB1/SRC-3* gene (amplified in breast cancer gene 1, a steroid receptor coactivator and an AR coactivator) with prostate cancer risk in a multidisciplinary population-based case-control study in China was

5 evaluated. Genomic DNA of 189 prostate cancer patients and 301 healthy controls was used for the PCR-based assay. The *AIB1/SRC-3* CAG/CAA repeat length for all alleles ranged from 24 to 32, with the most common repeat length being 29. Homozygous 29/29 and heterozygous 28/29 were the most common genotypes, with 44% and 30% of the controls harboring these genotypes, respectively. Relative to
10 subjects homozygous for ≥ 29 CAG repeats ($\geq 29/\geq 29$ genotype), individuals carrying one shorter allele ($\geq 29/<29$ genotype) had a 26% increased risk (OR = 1.26, 95% CI = 0.85-1.86), while those homozygous for the shorter allele ($<29/<29$ genotype) had a 73% excess risk (OR = 1.73, 95% CI = 0.96-3.11). The combined effect of CAG repeat lengths in the *AR* and *AIB1/SRC-3* genes was also evaluated. Relative to men
15 with both the $\geq 29/\geq 29$ genotype of the *AIB1/SRC-3* gene and a long CAG repeat length (≥ 23) in the *AR* gene, those with both the $<29/<29$ *AIB1/SRC-3* genotype and a short CAG repeat length in the *AR* gene (<23) had a 2.9-fold risk (OR = 2.86, 95% CI = 1.28-6.40). A similar was seen for the combined effects of the *AIB1/SRC-3* marker with the GGN marker of the *AR* gene. Together, these data indicate that the
20 CAG/CAA repeat length in the *AIB1/SRC-3* gene is associated with prostate cancer risk in Chinese men and that the combination of CAG/CAA repeat lengths in both the *AIB1/SRC-3* and *AR* genes provide a useful marker for clinically significant prostate cancer.

a) Materials and methods

25

(1) Study subjects

Details of the study have been described previously (9,16-18 AIB1) which are herein are incorporated by reference for at least material related to the epidemiological study. Cases of primary prostate cancer (ICD9 185) newly diagnosed between 1993 and 1995 were identified through a rapid reporting system
30 established between the Shanghai Cancer Institute and 16 collaborating hospitals in urban Shanghai. Cases were permanent residents in 10 urban districts of Shanghai (henceforth referred to as Shanghai) who had no history of other cancer. Contrary to many Western countries, prostate cancer screening is not widespread in China; therefore cases in this study were clinically significant prostate cancers who

5 presented with symptoms.

Based on the personal registry cards of all adults over age 18 residing in urban Shanghai (maintained at the Shanghai Resident Registry), male population controls were selected randomly from the 6.5 million permanent residents of Shanghai and frequency-matched to the expected age distribution (5-year category)
10 of prostate cancer cases. Included controls were negative for prostate cancer based on digital rectal exam and trans-rectal ultrasound.

Information on potential risk factors was elicited through an in-person interview by trained interviewers using a structured questionnaire. The interview included information on demographic characteristics; dietary and smoking history;
15 consumption of alcohol and other beverages; medical history; family history of cancer; physical activity; body size; and sexual behavior. Of the 268 eligible cases (95% of the cases diagnosed in Shanghai during the study period), 243 (91%) were interviewed. After a consensus review by both the Chinese and American pathologists, four cases were classified as having benign prostatic hyperplasia and
20 excluded from the study. Of the 495 eligible controls, 472 (95%) were interviewed. Most non-responses were due to refusal.

b) Blood collection and DNA extraction

Two hundred cases (82% of those interviewed) and 330 controls (70%) provided 20 ml of fasting blood for the study. The blood samples were processed at
25 a central laboratory in Shanghai. The buffy coat samples were first stored at -70°C and then shipped to the U.S. in dry ice for DNA extraction at the American Type Culture Collection (Manassas, VA), using a standard DNA extraction protocol. Quality control procedures showed no evidence of contamination, and DNA purity and length were satisfactory. After DNA extraction, 190 cases and 305 controls had
30 sufficient DNA for genotyping. DNA samples were arranged in case-control pairs/triplets to minimize day-to-day laboratory variation, and laboratory personnel were masked to case-control status.

5 c) Genotyping

The *AIB1/SRC-3* gene. The polyglutamine region of the *AIB1/SRC-3* protein is encoded by two glutamine codons in the *AIB1/SRC-3* gene on chromosome 20 (GenBank accession number AF012108): CAG and CAA. The usual sense codon sequence of the polyglutamine stretch is (CAG)_n CAA (CAG)_n (CAA CAG)₄ CAG CAA (CAG)₂ CAA SEQ ID NO:5. The two variable-length tracks of CAG repeats ((CAG)_n) usually contain six repeats between nucleotides 3930 and 3947, and nine repeats between nucleotides 3951 and 3977, for a total repeat length of 29 (19 *AIB1*) (which is herein incorporated by reference for material related to the CAG repeat.) This polymorphism has previously been described by Shirazi et al., though while Shirazi et al. scored genotypes of this marker using only the two variable (CAG)_n stretches (19 *AIB1*), herein the data was scored as the total number of continuous CAG and CAA triplets in the entire polyglutamine region of the *AIB1/SRC-3* gene, as has been done more recently (20,21 *AIB1*) (both of which are herein incorporated by reference for material related to the CAG and CAA repeats).

The number of CAG/CAA repeats in the polyglutamine stretch of the *AIB1/SRC-3* gene was determined by amplifying the gene's C-terminal polyglutamine region in each sample using custom flanking primers (5'-TCATCACCTCCGACAACAGAGG-3' and 5' (SEQ ID NO:1)-
25 TATGGAAACTGTTGCGGAGGAG-3' (SEQ ID NO:2) and the Advantage 2 Polymerase System (Clontech). The number of CAG/CAA repeats was determined by electrophoresis of the PCR products on an acrylamide gel and comparison to molecular weight standards. For confirmation, PCR products from selected samples were subsequently purified, using the PCR Product Purification Kit (Qiagen), and
30 sequenced directly, using the Big Dye Terminator Cycle Sequencing Ready Reaction Kit (PE Biosystems).

The AR gene. Genotypes of both the CAG (polyglutamine) and GGN (polyglycine) repeat length polymorphisms in exon 1 of the AR gene (located on the X chromosome) were determined as described previously (22 AIB1) (which is

5 herein incorporated by reference for material related to the AR gene). Briefly, two
sets of oligonucleotide primers flanking each of the two polymorphic regions for use
in DNA amplification and direct sequencing were designed. For the polyglutamine
stretch, the number of continuous CAG or CAA triplets was counted directly, while
for the polyglycine stretch, the number of continuous GGN repeats (where N
10 represents T, G, or C) was counted directly

Quality control. Because the PCR procedure is prone to contamination, a
negative water-blank control was always included in each batch of the PCR
reactions (usually 9-18 DNA samples plus one negative control). If the negative
control was shown to be positive, the assay was repeated for the entire batch.
15 Twenty-four split samples from a single individual were spaced at intervals among
the study samples to assess the reproducibility of genotyping. For the *AIB1/SRC-3*
gene, all of the 24 split samples had a CAG/CAA repeat length of 29 in both alleles.
Of the 21 split samples with AR CAG results, 19 (90%) had the same repeat number
(repeat length=23); one had one more, and one had one less repeat. Of the 20
20 samples with AR GGN results, 19 (95%) had the same repeat number (repeat
length=23) and one had one less repeat.

Statistical Analysis. Unconditional logistic regression models were used to
derive odds ratios (ORs) and corresponding 95% confidence intervals (CIs) to
estimate the prostate cancer risks associated with *AIB1/SRC-3* genotypes (23 AIB1)
25 (which is herein incorporated by reference for material related to statistical analysis).
The distribution of the number of the CAG/CAA repeat lengths among controls
were used to derive the median cutoffs used to calculate the ORs. Because the
AIB1/SRC-3 gene is located on chromosome 20, each individual carries two alleles.
In contrast, since the AR gene is located on the X chromosome, there is only one
30 allele for each individual. AR CAG and the AR GGN polymorphisms in this Chinese
population (9 AIB1), subjects in the current *AIB1/SRC-3* analysis were grouped by
<23 versus ≥23 repeats in analyses stratified by each of the two AR gene
polymorphisms. The level of significance for all results reported herein is 0.05.

5

d) Results

Age at diagnosis ranged from 50 to 94 (median 73) for cancer cases. Due to the lack of widespread prostate cancer screening in China, cases in this study were mostly men with clinically significant prostate cancer. Accordingly, about two-thirds of the cases were diagnosed as having advanced (regional/remote stages) cancer, and most tumors were moderately or poorly differentiated. Most cases were symptomatic at diagnosis, and 77% had serum prostatic specific antigen levels greater than 10 ng/ml (median 87 ng/ml). Compared to population controls, cases had significantly higher caloric intake; had significantly larger waist-to-hip ratios; and were somewhat less likely to be married, have attended college, or be smokers or drinkers, though not significantly so (data not shown).

The distribution of the alleles and genotypes of the *AIB1/SRC-3* gene CAG/CAA repeat length marker by case-control status is shown in Table 1. Among controls, the CAG/CAA repeat length for all alleles ranged from 24 to 32, with 29, 28, and 26 being the most common repeat lengths (65.6, 23.5, and 6.5%, respectively). Eighty-eight percent of the controls had at least one 29 allele. Homozygous 29/29 and heterozygous 28/29 were the most common genotypes, with 44% and 30% of the controls harboring these genotypes, respectively. The observed genotype frequencies were in close agreement with those predicted from the allele frequencies under Hardy-Weinberg equilibrium.

Table 1. Frequencies of *AIB1/SRC-3* CAG/CAA repeat length alleles and genotypes in prostate cancer cases and controls, China

<i>AIB1/SRC-3</i> allele	Cases		Controls	
	n	%	n	%
24	0	0.0	1	0.2
26	22	5.8	39	6.5
27	6	1.6	7	1.2
28	114	30.0	140	23.5
29	224	58.9	391	65.6
30	11	2.9	13	2.2
31	1	0.3	5	0.8

	36			
32	2	0.5	0	0.0
Total	380		596	
<i>AIB1/SRC-3</i> genotype	n	%	n	%
24/29	0	0.0	1	0.3
26/26	0	0.0	1	0.3
26/27	0	0.0	1	0.3
26/28	5	2.6	6	2.0
26/29	17	8.9	26	8.7
26/30	0	0.0	2	0.7
26/31	0	0.0	2	0.7
27/28	0	0.0	1	0.3
27/29	5	2.6	5	1.7
27/30	1	0.5	0	0.0
28/28	23	12.1	21	7.0
28/29	59	31.1	88	29.5
28/30	3	1.6	1	0.3
28/31	1	0.5	2	0.7
29/29	67	35.3	130	43.6
29/30	7	3.7	10	3.4
29/31	0	0.0	1	0.3
29/32	2	1.1	0	0.0
Total	190		298	

5

Relative to men homozygous for 29 CAG/CAA repeats in the *AIB1/SRC-3* gene (29/29 genotype), subjects homozygous for the 28 allele had a significant risk increase (OR = 2.12, 95% CI = 1.09-4.12, Table 2). Subjects with the 28/29 genotype had a non-significant increased risk relative to the 29/29 genotype (OR = 1.30, 95% CI = 0.83-2.03), as did men with one 29 allele and one 24, 26, or 27 allele (OR = 1.33, 95% CI = 0.72-2.47) and men with one 29 allele and one 30, 31, or 32 allele (OR = 1.58, 95% CI = 0.62-4.01).

Table 2. Age-adjusted ORs for prostate cancer in relation to CAG/CAA repeat lengths in the *AIB1/SRC-3* gene, China

15

Allele 1	Allele 2	Cases	Control	OR ^a	95% CI
29	29	67	130	1.00	-
28	29	59	88	1.30	0.83-2.03
24,26,27	29	22	32	1.33	0.72-2.47
30,31,32	29	9	11	1.58	0.62-4.01
28	28	23	21	2.12	1.09-4.12
26,27,28	30,31	5	7	1.38	0.42-4.52

			37		
26,27	26,27,28	5	9	1.08	0.35-3.35
≥29	≥29	76	141	1.00	-
≥29	<29	86	127	1.26	0.85-1.86
<29	<29	28	30	1.73	0.96-3.11

5

Based on the median CAG/CAA repeat length of 29, the various *AIB1/SRC-3* alleles were collapsed into those with 29 or more repeats (≥29) and those with less than 29 repeats (<29). Relative to men with the ≥29/≥29 genotype, men with one long and one short allele (≥29/<29 genotype) had a moderate but non-significant risk elevation (OR = 1.26, 95% CI = 0.85-1.86). Those with two short alleles (<29/<29 genotype) had a non-significant 73% increased risk (OR = 1.73, 95% CI = 0.96-3.11) relative to men with two long alleles (≥29/≥29 genotype).

Risks of prostate cancer associated with various repeat lengths in both the *AIB1/SRC-3* gene CAG/CAA polymorphism as well as the two polymorphisms of the *AR* gene are shown in Table 3. Relative to men both homozygous for the long CAG/CAA allele (≥29/≥29 genotype) of the *AIB1/SRC-3* gene and having a long *AR* CAG repeat length (≥23 repeats), men both homozygous for the short *AIB1/SRC-3* CAG/CAA allele (<29/<29) and having a short *AR* CAG repeat length (<23) had a significant 2.9-fold risk (OR=2.86, 95% = 1.28-6.40). Similarly, those men both homozygous for the short *AIB1/SRC-3* CAG/CAA allele and having a short GGN repeat length (<23) in the *AR* gene had a non-significant 2.3-fold risk (OR=2.29, 95% CI = 0.73-7.15) relative to men both homozygous for long *AIB1/SRC-3* CAG/CAA allele (≥29/≥29 genotype) and having a long *AR* GGN repeat length (≥23 repeats).

5 Table 3. ORs ^a for prostate cancer in relation to *AIB1/SRC-3* CAG/CAA and *AR* CAG or GGN repeat lengths, China

AR polymorphisms	CAG/CAA repeat length genotypes of the <i>AIB1/SRC-3</i> gene					
	$\geq 29/\geq 29$		$\geq 29/< 29$		$< 29/< 29$	
	n1/n2 ^b	OR	95% CI	n1/n2 ^b	OR	95% CI
CAG repeat length ^c						
≥ 23	30/68	1.00	-	35/69	1.15	0.63-2.08
< 23	45/71	1.43	0.81-2.54	51/58	1.99	1.12-3.53
GGN repeat length ^c						
≥ 23	57/11	1.00	-	72/99	1.42	0.92-2.21
< 23	17/26	1.28	0.64-2.56	14/24	1.15	0.55-2.40

^a Adjusted for age.

^b n1=number of cases, n2=number of controls.

10 ^c Median number of repeats among controls was used for the cutoff.

There was no correlation between the repeat lengths in the *AR* and *AIB1/SRC-3* genes. In addition, the number of CAG/CAA repeats in the *AIB1/SRC-3* gene did not correlate with education, body mass index, waist-to-hip ratio, total caloric intake, serum levels of sex hormones (testosterone; DHT; 5 α -androstane-
 15 3 α ,17 β -diol glucuronide; and estradiol), and sex hormone binding globulin. These variables therefore were not included in the logistic model for adjustment. In addition, odds ratios were materially unchanged when the analysis was stratified by clinical stage (localized versus advanced stage disease, data not shown).

Results from this population-based case-control study in China indicate that
 20 men with fewer than 29 CAG/CAA repeats in the *AIB1/SRC-3* alleles have an increased risk of clinically significant prostate cancer. Furthermore, our results suggest that this effect, though independent of *AR* genotypes, is more pronounced among men with a smaller number of *AR* CAG repeats.

The observed association with CAG/CAA repeat length in the *AIB1/SRC-3*
 25 gene is biologically plausible. Data from transient transfection studies show that the *AIB1/SRC-3* coactivator enhances *AR* transcriptional activity in the presence of

5 DHT (12 AIB1), suggesting that the AIB1/SRC-3 coactivator, in conjunction with AR, may increase androgenic activity within the prostate gland. Amplification of the AIB1/SRC-3 gene has been implicated in the etiology of several other hormone-dependent cancers as well, including breast and ovarian cancers (24 AIB1). Furthermore, recent clinical data suggest that overexpression of AR in prostate
10 tumors may contribute to hormone sensitivity and tumor progression (25 AIB1).

Racial/ethnic variation in the AIB1/SRC-3 CAG/CAA repeat length mirrors the risk patterns of prostate cancer in high- and low-risk populations (19,26 AIB1), thus indirectly supporting a role of AIB1/SRC-3 in prostate cancer etiology. In a small survey of 112 African Americans, 19 Chinese, and 18 Caucasians, the allele
15 frequency of 29 CAG/CAA repeats was 61%, 76%, and 58%, respectively.

Chinese men have a longer mean CAG repeat length in the AR gene than Western men, and that a shorter AR CAG repeat length was associated with an increased risk of prostate cancer in this low-risk population (9 AIB1). The observed association with CAG/CAA repeat length in the AIB1/SRC-3 gene is independent of
20 the AR gene: regardless of CAG repeat length in the AR gene, men homozygous for the shorter AIB1/SRC-3 allele (the $<29/<29$ genotype) had a higher risk than those homozygous for the long allele ($\geq 29/\geq 29$ genotype). However, the risk associated with the homozygous $<29/<29$ AIB1/SRC-3 genotype was more pronounced among those with the short AR CAG repeat length.

25 **2. Example 2. Relationship of CAG length in the androgen receptor**

The length of the polymorphic CAG trinucleotide repeat in the polyglutamine region of the androgen receptor (AR) gene is inversely correlated with the transactivation function of the AR. A population-based case-control assay
30 in China addressed CAG and other polymorphisms of the AR gene and their association with clinically significant prostate cancer in this low-risk population. Genomic DNA from 190 prostate cancer patients and 304 healthy controls were used for direct sequencing to evaluate the relationship of CAG and GGN (polyglycine) repeat length in the AR gene. Relative to western men, the subjects had a longer

- 5 CAG repeat length, with a median of 23 and only 10% of the subjects having a CAG repeat length shorter than 20. Men with a CAG repeat length shorter than 23 (median length) had a 65% increased risk of prostate cancer (OR=1.65, 95% CI 1.14-2.39), compared to men with a CAG repeat length of 23 or longer.

- For the GGN tract (GGT₃GGG₁GGT₂GGC_n), based on the sequencing results
10 from 481 samples, it is shown that even though GGC regions in the polyglycine tract are highly variable, there are no mutations or polymorphisms in the GGT and GGG regions. Seventy two percent of the subjects had a GGN repeat length of 23, and those with a GGN repeat length shorter than 23 had a 12% increased risk of prostate cancer (95% CI 0.71-1.78), compared to those with 23 or more GGN repeats. This
15 not only confirms that Chinese men do have a longer CAG repeat length than western men, but also represents the first population assay to show that even in a very low-risk population, a shorter CAG repeat length confers a higher risk of clinically significant prostate cancer. These results indicate that CAG repeat length can serve as a useful marker to identify a subset of individuals at higher risk of
20 developing clinically significant prostate cancer.

a) Materials and methods

- Subjects.** Details of the assay have been described previously (28AR which is herein incorporated by reference for material related to the assay). Briefly, cases of primary prostate cancer (ICD9 185) newly diagnosed between 1993 and 1995 were
25 identified through a rapid reporting system established between the Shanghai Cancer Institute (SCI) and 28 collaborating hospitals in urban Shanghai. Cases were permanent residents in 10 urban districts of Shanghai (henceforth referred to as Shanghai) who had no history of any other cancer. Of the 268 eligible cases (representing 95% of the cases diagnosed in urban Shanghai during the study
30 period), 243 (91%) were interviewed in person by trained interviewers. Four of the cases were later classified as having benign prostatic hyperplasia and excluded from the study after a consensus review by both Chinese and U.S. pathologists.

Based on the records at the Shanghai Resident Registry, which contains personal identification cards for all adult residents over age 18 in urban Shanghai,

5 healthy subjects who were free of cancer were selected randomly from permanent residents of Shanghai (6.5 million), frequency-matched to the expected age distribution (5-year category) of prostate cancer cases. Of the 495 eligible controls without a history of cancer, 472 (95%) were interviewed.

Information on potential risk factors was elicited through an in-person
10 interview by trained interviewers using a structured questionnaire. The interview included information on demographic characteristics; dietary history; smoking history; consumption of alcohol and other beverages; medical history; family history of cancer; physical activity; body size; and sexual behavior.

Blood collection and DNA extraction. Two hundred cases (84% of those
15 interviewed) and 330 controls (70%) provided 20 ml of fasting blood for the study. The blood samples were processed within three hours of collection at a central laboratory in Shanghai and stored at -70°C. The frozen buffy coat samples (separated from 5 ml of blood) were later shipped to the U.S. on dry ice for DNA
20 extraction at the American Type Culture Collection (Manassas, Virginia) with standard protocols. DNA purity, yield, and length were satisfactory and there was no evidence of DNA degradation or RNA contamination. Following DNA
25 extraction, 191 cases and 304 controls had sufficient DNA for AR genotyping at the University of Rochester. DNA samples for cases and controls were grouped into pairs to minimize the effect of day-to-day laboratory variation. Laboratory personnel were blinded to the case-control status.

Molecular analysis and assessment of the CAG and GGN repeats. As part of an ongoing molecular analysis of the AR gene, genomic DNA from the 495 subjects was used to determine the usual sense codon sequence and the exact number of CAG and GGN repeats in exon 1 of the AR gene through PCR
30 amplification and DNA sequencing. For the CAG repeat analysis, a set of oligonucleotide primers that flank the CAG repeat, 5'-GCTCTGGGACGCAACCTCTCT-3' (SEQ ID NO:7) and 5'-GCAGCGACTACCGCATCATCA-3' (SEQ ID NO:8), were designed for PCR amplification. A pair of nested primers, 5'-CGGG-GTAAGGGAAGTAGGTGGAAG-3' (SEQ ID NO:15), and 5'-

5 CTCTACGATGGGCTTGGGGAGAAC-3' (SEQ ID NO:16) was selected for DNA sequencing. For GGN analysis, the oligonucleotide primers 5'-ACCCTCAGCCGCGCTTCCTCATC-3' (SEQ ID NO:11) and 5'-CTGGGATAGGGCACTCTGCTCAAC-3' (SEQ ID NO:12) were used for both PCR amplification and sequencing. The PCR products of the CAG and GGN
10 repeats were amplified, using the Advantage 2 Polymerase System (Clontech) and the Advantage-GC Genomic Polymerase System (Clontech), respectively. Subsequently, these PCR products were purified, using the PCR Product Purification Kit (Qiagen), and sequenced directly, using the Big Dye Terminator Cycle Sequencing Ready Reaction Kit (PE Biosystems). All reactions were optimized to
15 reach consistent results, using genomic DNA samples extracted from cell lines. For the polyglutamine tract ((CAG)_nCAA), the number of CAG triplets was counted to yield the length of CAG repeats. For the polyglycine tract (GGT₃GGG₁GGT₂GGC_n) (SEQ ID NO:17), the usual sense codon sequence of the GGN tract is: three GGT, one GGG, two GGT, followed by a variable number of GGC repeats. For example,
20 a GGN repeat length of 23 in our study corresponded to a PCR fragment of 217 bp, encompassing 3 GGT, 1 GGG, 2 GGT, and 17 GGC triplets.

Because the PCR procedure is prone to contamination, a negative control (water blank) was always included in each batch of PCR reactions (usually 9-18 samples plus one negative control). The assay for one batch (9 samples) was
25 repeated with new reagents because of an indication of minor contamination. Because exon 1 of the AR gene is GC-rich with CAG and GGN repeats, this region is difficult to amplify. Several samples had to be amplified and sequenced more than once. Overall, five (1%) of the 495 samples could not be typed for CAG repeats due to insufficient DNA or sequencing problems, while 14 (2.8%) could not
30 be typed for GGN repeats for similar reasons. The percentages of samples that were unsuccessfully genotyped were similar in cases and controls.

Twenty-four split samples from the same individual were included as quality control samples to assess the reproducibility of genotyping. Of the 24 quality control samples, 23 and 20 were amplified and sequenced successfully for the CAG

5 and GGN repeats, respectively. Of the 23 samples with CAG results, 21 (91%) had the same repeat length of 23, one had 24, and one had 22. Of the 20 samples with GGN results, 19 (95%) had the same repeat length of 23 and the other had length 24.

Statistical Analysis. The mean numbers of CAG and GGN repeats were compared in cases and controls using the t test. Unconditional logistic regression
10 models were used to estimate odds ratios (ORs) and their corresponding 95 percent confidence intervals (CIs) for prostate cancer in relation to CAG and GGN repeat lengths (29AR). Repeat lengths were examined first as continuous variables and later as categorical variables. The distributions of the number of CAG or GGN repeats among controls were used to derive the median or tertile cutoffs used to
15 calculate ORs. In addition, the combined effects of CAG and GGN were evaluated based on the median lengths within the controls. The relationships between age, CAG and GGN repeat length, and other variables were assessed by Spearman correlation and analysis of variance.

b) Results

20 Selected characteristics of cases and controls are shown in Table 4. Compared to controls, cases had higher caloric intake and higher levels of education and waist-to-hip ratio and were less likely to use cigarettes or alcohol. Age at diagnosis ranged from 50 to 94 (median 73) for cancer cases. Sixty-nine cases (36%) were diagnosed as having localized cancer, and most tumors (72%) were
25 moderately or poorly differentiated.

Table 4 Selected characteristics of prostate cancer cases and population controls, China

<u>Characteristics</u>	<u>Cases (n=191)</u>		<u>Controls (n=304)</u>	
	<u>Mean</u>	<u>(S.D.)</u>	<u>Mean</u>	<u>(S.D.)</u>
Age (yrs)	72.2	(7.7)	71.9	(7.3)
Total calories (Kcal/day)	2457.0	(647)	2342.0	(731)
Height (cm)	167.9	(6.0)	167.6	(5.8)
Weight (Kg)	61.3	(8.4)	61.5	(10.1)
BMI (Kg/m ²)	21.8	(2.9)	21.9	(3.3)

	44	
Waist circumference (cm)	82.6 (10.4)	82.5 (10.7)
Hip circumference (cm)	90.7 (8.9)	92.6 (8.5)
Waist-to-hip ratio	0.91 (0.05)	0.89 (0.05)
% married	89.5	92.1
% with education greater than high school	34.6	25.3
% smokers	56.5	65.1
% alcohol users	31.4	42.1
Clinical stage (%)		
Localized	36.3	
Regional	30.5	
Remote	32.1	
Histologic grade (%)		
Well differentiated	7.9	
Moderately differentiated	31.0	
Poorly differentiated	41.0	
Cannot be assessed	20.1	

5

Because the AR gene is located on the X chromosome, only one copy of the gene is present in men. For the polyglutamine tract (CAG_nCAA), there was no variation in the CAA sequence among the 490 samples analyzed. The number of CAG repeats ranged from 10 to 34. About 65% of the study subjects had a CAG repeat length that ranged from 21 to 24, but only 1% of the subjects had a CAG length longer than 30 repeats (Table 5).

10

Table 5 *Distribution of number of CAG repeats in the androgen receptor gene in prostate cancer cases and controls, China*

Cases (n=190)			Controls (n=300)	
<u>No. of CAG repeats</u>	<u>N</u>	<u>%</u>	<u>N</u>	<u>%</u>
10	0	0.0	1	0.3
14	0	0.0	1	0.3

15

		45		
15	4	2.1	2	0.7
16	3	1.6	2	0.7
17	1	0.5	3	1.0
18	7	3.7	9	3.0
19	7	3.7	12	4.0
20	13	6.8	17	5.7
21	24	12.6	32	10.7
22	57	30.0	67	22.3
23	22	11.6	46	15.3
24	21	11.1	48	16.0
25	16	8.4	19	6.3
26	9	4.7	17	5.7
27	3	1.6	15	5.0
28	2	1.1	2	0.7
29	0	0.0	3	1.0
30	0	0.0	1	0.3
31	0	0.0	2	0.7
32	0	0.0	0	0.0
33	0	0.0	1	0.3
34	1	0.5	0	0.0
Median	22		23	

5

Although the median number of CAG repeats in controls was only slightly larger than that in cases (23.0 vs. 22.0), there was a shift toward longer repeat length among controls (Figure 1). For CAG repeat length shorter than 23, cases had higher percentages than controls in 6 of the 10 categories. However, for CAG repeat length longer than 22, controls had higher percentage than cases in 8 of the 12 categories. Age at diagnosis and stage of cancer were not related to CAG repeat length, with similar distribution and average number of CAG and GGN repeat lengths in various age categories and three clinical stages.

For the polyglycine tract (GGT₃GGG₁GGT₂GGC_n) (SEQ ID NO:17), there

- 5 was no variation in the codon usage or the number of GGT and GGG trinucleotides in all of the 481 samples analyzed, although the number of GGC repeats was highly variable. The pattern was always three GGT, one GGG, two GGT, followed by a variable number of GGC. The number of GGN repeats among study subjects ranged from 15 to 27 (the number of GGC repeats thus ranged from 9 to 21) (Table 6).
- 10 About 72% of the study subjects had a GGN repeat length of 23.

Table 6 *Distribution of number of GGN repeats in the androgen receptor gene in prostate cancer cases and controls, China*

<u>No. of GGN repeats</u>	<u>Cases (n=187)</u>		<u>Controls (n=295)</u>	
	<u>N</u>	<u>%</u>	<u>N</u>	<u>%</u>
14	0	0.0	1	0.3
15	1	0.5	1	0.3
16	2	1.1	1	0.3
17	2	1.1	0	0.0
18	0	0.0	1	0.3
19	19	10.2	20	6.8
20	2	1.1	2	0.7
21	3	1.6	0	0.0
22	10	5.3	24	8.2
23	136	72.7	212	72.1
24	10	5.3	24	8.2
25	2	1.1	1	0.3
27	0	0.0	1	0.3
Median	23		23	

- 15 Risks of prostate cancer associated with CAG and GGN repeat lengths are shown in Table 7. When the number of CAG repeats was included in the model as a continuous variable, there was a 7% increase in the risk of prostate cancer for each decrement in length of one CAG repeat (OR=1.07, 95% CI 1.00-1.15). The risks associated with decrements of three and six repeats were 1.21 (95% 1.14-1.32) and

- 5 1.42 (95% CI 1.22-1.61), respectively. When the median repeat length was used to dichotomize study subjects, men with a CAG repeat length shorter than 23 had a 65% increased risk (OR=1.65, 95% CI 1.14-2.39), compared to men with a CAG repeat length of 23 or longer. Relative to the highest tertile of CAG repeat length (≥ 24), men in the second and first tertiles (22-23 and <22) had ORs of 1.45 and 1.55, respectively ($P_{\text{trend}}=0.06$).
- 10

Table 7 Odds ratios (ORs)^a and 95% confidence intervals (CIs) for prostate cancer in relation to the number of CAG and GGN repeats in the androgen receptor gene, China

	Cases	Controls		
<u>Number of CAG and GGN repeats</u>	<u>No.</u>	<u>No.</u>	<u>ORa</u>	<u>95% CI</u>
No. of CAG repeats				
Continuous (per decrement of one CAG repeat)	190	300	1.07	1.00-1.15
Median				
≥23	74	154	1.00	-
<23	116	146	1.65	1.14-2.39
Tertile				
≥24	52	108	1.00	-
22-23	79	113	1.45	0.93-2.25
<22	59	79	1.55	0.96-2.49
			Linear trend p=0.06	
No. of GGN repeats				
Continuous (per decrement of one GGN repeat)	187	294	1.07	0.96-1.20
Median				
≥23	147	239	1.00	1.00
<23	39	56	1.12	0.71-1.78
Combined number of CAG and GGN repeats				
CAG>23, GGN ≥23	53	120	1.00	-

	48			
CAG \geq 23, GGN < 23	19	29	1.48	0.76- 2.88
CAG<23, GGN \geq 23	94	115	1.85	1.21- 2.82
CAG<23, GGN < 23	20	26	1.75	0.90- 3.41

5 ^aAdjusted for age (continuous).

Similarly, men with a shorter GGN repeat length had a higher risk of prostate cancer. Each decrement of one GGN repeat length was associated with a 7% increase in risk (OR= 1.07, 95% CI 0.96-1.20). Men with a GGN repeat length shorter than the median length of 23 had a 12% increase in prostate cancer risk, compared to those with 23 or more repeats. Because more than 72% of the subjects had 23 GGN repeats, the ORs by tertiles for GGN repeats were not estimated.

Also shown in Table 7 are the ORs associated with combined categories of CAG and GGN repeat lengths. Men with both CAG and GGN repeat lengths shorter than 23 had a 75% elevated risk of prostate cancer. There was little correlation between the number of CAG and GGN repeats ($r=-0.03$, $p>0.05$).

The number of CAG or GGN repeats did not correlate with age, education, body mass index, waist-to-hip ratio, total calories, smoking, or drinking. These variables therefore were not included in the model for adjustment. The ORs were materially unchanged after further adjustment for benign prostatic hyperplasia (BPH), although the cases had a higher prevalence of BPH (57% vs. 23%) and there was a non-significant moderate association between CAG or GGN repeat lengths and BPH (data are reported separately). Associations of CAG or GGN repeat length were similar across all stages of disease at diagnosis (data not shown).

These results confirm that a shorter CAG repeat length is associated with an increased risk of clinically significant prostate cancer. A shorter length of GGN repeat also appears to increase the risk of prostate cancer, but the magnitude of excess risk was smaller.

The observed inverse association with AR polymorphisms is biologically

5 plausible, as laboratory studies have shown that a long polyglutamine chain (>30 repeats) in the AR gene is associated with androgen insensitivity and reduced AR transactivation activity (13,14AR). *In vitro* transfection studies also have demonstrated that elimination of the polyglutamine tracts results in elevated transcriptional activities (9,11,16AR). Clinical studies have suggested that alteration
10 in the AR function, either through polymorphisms of CAG repeat length or somatic mutations, may be associated with tumor progression. For example, the progression from latent to clinically invasive prostate cancer is initially androgen-dependent, although some tumors later become androgen-independent (thus becoming non-responsive to hormonal treatment). Several non-germline-related changes of the AR
15 gene, including amplification of the AR gene (usually a key step in the transition from a hormone-sensitive to a hormone-refractory state in prostate tumors) (31,32AR), AR somatic mutations (identified throughout transactivation, DNA binding, and ligand binding domains) (33,34AR), and contraction of CAG repeat length in cancer cells (32AR), have been shown to be associated with tumor
20 aggressiveness, cancer progression, and failure of hormonal therapy. AR expression studies in the majority of prostate tumors, including those that have become refractory to hormonal therapy, also suggest that AR plays a key role in androgen-independent tumors (35,36AR).

The inverse relationship between CAG repeat length and AR transcriptional
25 activity (thus androgen sensitivity) is the currently recognized underlying molecular mechanism by which AR polymorphisms modulate prostate cancer risk. Because transcriptional activation of the AR gene is influenced by not only polymorphisms in the AR gene but also a number of other factors, including tissue levels of dihydrotestosterone (DHT), estradiol, insulin-like growth factors, and AR
30 coactivators (37-43AR), it is likely that these factors may also affect prostate cancer risk by mediating transcriptional activities. Several AR coactivators, including AR-associated proteins (ARA70, ARA55), AIB1 (Amplified in Breast Cancer -1), CBP (cyclic AMP responsive element binding protein), Rb, and BRCA1, have been shown to enhance AR-mediated transcriptional activity from 2- to 10-fold,

5 suggesting that *in vivo* coactivators are essential in attaining optimal AR
transactivation in response to androgens (40-43AR).

It has been suggested that variations in CAG repeat length in the AR gene
between populations may explain part of the large racial difference in prostate cancer
risk and that a shorter CAG repeat length reported for African Americans may
10 contribute to some of their higher risk of prostate cancer, although currently no data
are available from this population. Our results confirm that, relative to western men,
Chinese men do indeed have a longer CAG repeat length. For example, 22% of the
1,722 white men in two U.S. studies (17,18AR) had a CAG repeat length shorter
than 20 vs. only 10% in our study and 55% reported for African Americans in a
15 cross-sectional survey (26,27AR). Inverse associations have also been reported for
Caucasians, suggesting that the underlying biological mechanism in various racial
groups may be similar and that the polymorphisms of AR may be related, in part, to
racial difference in prostate cancer risk.

The common polymorphism of the AR gene confers variable risk upon all
20 individuals, which in turn may result in a much larger proportion of prostate cancer
cases attributable to having fewer CAG repeats. Assuming that the CAG
polymorphism association is causal, it is estimated that 25% (95% CI 9% to 41%) of
the cases in Shanghai can be attributed to a CAG repeat length shorter than 23.
Using the CAG repeat length distribution in the two U.S. studies among white men
25 (17,18AR), it was estimated that 3-7%% of cases in the U.S. white men can be
attributed to the CAG polymorphism (repeat length <23) and that this polymorphism
alone potentially accounts for at least 5% of the difference in incidence between
Chinese and U.S. men.

Similar to two previous studies (17,18AR), it was found that the number of
30 GGN repeats clusters around 23 (in the study of Stanford et al., only the number of
GGC repeats was counted and 15 was the peak of the repeat, which corresponds to
21 GGN repeats), and that a shorter GGN repeat length appears to be associated with
a moderate increase in prostate cancer risk. Twenty-three GGN repeats may
represent the coding sequence for optimal AR protein conformation and activity,

5 because over 70% of the study subjects in our study as well as in studies of western men had a GGN repeat length of 23.

Although it is well established that (GGC)_n repeats in the polyglycine tract (GGT₃GGG₁GGT₂GGC_n) (SEQ ID NO:17) of the AR gene is polymorphic, to date there has been little information on variations in the GGG and GGT regions of the polyglycine tract, because these regions are GC-rich and technically it has been difficult to amplify these regions. Our study represents the first successful effort to sequence the exact codon usage and number of the GGN trinucleotide repeats in a large number of population-based samples. It was showed that GGT and GGG regions were quite stable and there were no variations in these two regions in all of the 481 DNA samples analyzed.

F. AIB1 references

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G. AR References

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- 40

H. Sequences

- 45 SEQ ID NO:1 first primer
5'-TCA TCA CCT CCG ACA ACA GAG G-3'
- SEQ ID NO:2 second primer
5'-TAT GGA AAC TGT TGC GGA GGA G-3'

5

SEQ ID NO:3 complement to first primer
5'-CCT CTG TTG TCG GAG GTG ATG A-3'

10 SEQ ID NO:4 complement to second primer
5'-CTC CTC CGC AAC AGT TTC CAT A -3'

SEQ ID NO:5 general CAG/CAA sequence
5'-(CAG)_N CAA (CAG)_N (CAACAG)₄ CAG CAA CAG CAG CAA-3'
Nucleotides 1-3 and 6-9 as a triplets must be at least one triple but can be any
15 multiple of triplets.

SEQ ID NO:6 one particular 29 mer sequence
5'-CAG CAG CAG CAG CAG CAG CAA CAG CAG CAG CAG CAG CAG
CAG CAG CAA CAG CAA CAG CAA CAG CAA CAG CAG CAA CAG CAG
20 CAA-3'

SEQ ID NO:7 first androgen receptor CAG primer
5'-GCT CTG GGA CGC AACCTCTCT-3'

25 SEQ ID NO:8 second androgen receptor CAG primer
5'-GCA GCG ACT ACC GCA TCA TCA-3'

SEQ ID NO:9 complement to first androgen receptor CAG primer
5'-AGAGAGGTTGCGTCCAGAGC-3'

30 SEQ ID NO:10 complement to second androgen receptor CAG primer
5'-TGATGATGCGGTAGTCGCTGC-3'

SEQ ID NO:11 first androgen receptor GGN primer
35 5'-ACCCTCAGCCGCGCTTCCTCATC-3''

SEQ ID NO:12 second androgen receptor GGN primer
5'-CTGGGATAGGGCACTCTGCTCAAC-3'

40 SEQ ID NO:13 complement to first androgen receptor GGN primer
5'-GATGAGGAAGCGGCGGCTGAGGGT-3'

SEQ ID NO:14 complement to second androgen receptor GGN primer
5'-GTTGAGCAGAGTGCCCTATCCCAG-3'

45 SEQ ID NO:15
5'-CGGG-GTAAGGGAAGTAGGTGGAAG-3'

SEQ ID NO:16
50 and 5'-CTCTACGATGGGCTTGGGGAGAAC-3'

5

SEQ ID NO:17
GGT₃GGG₁GGT₂GGC_n

SEQ ID NO:18 AIB1 sequence Genbank accession number XM_030032

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61 tccgattta aagctgagct gcgaggaaaa tggcggcggg aggatcaaaa tacttgctgg
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SEQ ID NO:20

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15 YFHTQ

5 What is claimed is:

1. A method for assessing the risk of prostate cancer in a human subject comprising determining the length of the contiguous CAG or CAA repeats in both AIB1 gene alleles of the subject and assessing whether the length of the CAG or CAA repeats is less than, equal to, or greater than 29 repeats, a length less than or
10 greater than 29 repeats in both alleles indicating an increased risk of prostate cancer in the subject.
2. The method of claim 1, wherein determining the length of the repeats comprises amplifying a region of both AIB1 gene alleles comprising the contiguous CAG or CAA repeat.
- 15 3. The method of claim 2, wherein the amplification is by PCR that produces two PCR products.
4. The method of claim 3, further comprising analyzing the PCR products by chromatography.
5. The method of claim 4, wherein the chromatography is gel
20 electrophoresis.
6. The method of claim 2, wherein the sequence of the PCR products is determined.
7. The method of claim 3, wherein the PCR product is produced using a first AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:3
25 and a second AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:4.
8. The method of claim 7, wherein the first AIB1 primer has the sequence set forth in SEQ ID NO:1 and the second AIB1 primer has the sequence set forth in SEQ ID NO:2.
- 30 9. The method of claim 1, wherein determining how many CAG or CAA repeats there are comprises sequencing the CAG or CAA repeats.

5 10. A method for assessing the risk of prostate cancer in a human subject comprising determining the length of the contiguous CAG or CAA repeats in the AIB1 gene alleles of the subject and assessing whether the length of the CAG or CAA repeats in each allele is less than, equal to, or greater than 29 repeats, and
10 determining the length of the contiguous CAG or CAA repeats in the androgen receptor gene of the subject and assessing whether the length of the CAG or CAA repeats is less than, equal to, or greater than 23 repeats, a length in at least one allele less than or greater than 29 repeats in the AIB1 gene and less than or greater than 23 repeats in the androgen receptor gene indicating an increased risk of prostate cancer in the subject.

15 11. The method of claim 10, wherein determining the length of the contiguous CAG or CAA repeats in the AIB1 gene alleles comprises amplifying a region of the AIB1 gene alleles comprising the CAG or CAA repeats and wherein determining the length of the CAG or CAA repeats in the androgen receptor gene comprises amplifying a region of the androgen receptor gene comprising the CAG or
20 CAA repeats.

 12. The method of claim 11, wherein the amplification of the regions of the AIB1 gene and the androgen receptor gene is by PCR that produces a first and a second AIB1 PCR product and an androgen receptor PCR product.

 13. The method of claim 12, further comprising analyzing the PCR products
25 by chromatography.

 14. The method of claim 13, wherein the chromatography is gel electrophoresis.

 15. The method of claim 12, wherein the sequence of the PCR products is determined.

30 16. The method of claim 12, wherein the AIB1 PCR product is produced using a first AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:3 and a second AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:4.

5 17. The method of claim 16, wherein the first AIB1 primer has the sequence set forth in SEQ ID NO:1 and the second AIB1 primer has the sequence set forth in SEQ ID NO:2.

10 18. The method of claim 12, wherein the androgen receptor PCR product is produced using a first androgen receptor CAG primer that selectively hybridizes with the sequence set forth in SEQ ID NO:9 and a second androgen receptor CAG primer that selectively hybridizes with the sequence set forth in SEQ ID NO:10.

19. The method of claim 18, wherein the first androgen receptor CAG primer has the sequence set forth in SEQ ID NO:7 and the second androgen receptor CAG primer has the sequence set forth in SEQ ID NO:8.

15 20. The method of claim 12, wherein the AIB1 PCR product is produced using a first AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:3 and a second AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:4. and wherein the androgen receptor PCR product is produced using a first androgen receptor CAG primer that selectively hybridizes with the sequence set forth in SEQ ID NO:9 and a second androgen receptor CAG primer that selectively hybridizes with the sequence set forth in SEQ ID NO:10.

20 21. The method of claim 20, wherein the first AIB1 primer has the sequence set forth in SEQ ID NO:1 and the second AIB1 primer has the sequence set forth in SEQ ID NO:2 and wherein the first androgen receptor CAG primer has the sequence set forth in SEQ ID NO:7 and the second androgen receptor CAG primer has the sequence set forth in SEQ ID NO:8.

22. The method of claim 10, wherein more than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and more than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

23. The method of claim 10, wherein more than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and less than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person

5 indicates an increased risk of prostate cancer in the subject.

24. The method of claim 10, wherein less than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and more than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

10 25. The method of claim 10, wherein less than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and less than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

15 26. The method of claim 10, wherein more or less than 29 contiguous CAG or CAA repeats in both alleles of the AIB1 gene of the person and more than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

20 27. The method of claim 10, wherein more or less than 29 contiguous CAG or CAA repeats in both alleles of the AIB1 gene of the person and less than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

25 28. The method of claim 10, wherein more than 29 contiguous CAG or CAA repeats in both alleles of the AIB1 gene of the person and more than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

29. The method of claim 10, wherein less than 29 contiguous CAG or CAA repeats in both alleles of the AIB1 gene of the person and less than 23 contiguous CAG or CAA repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

30 30. A method for assessing the risk of prostate cancer in a human subject comprising determining the length of the contiguous CAG or CAA repeats in the AIB1 gene alleles of the subject and assessing whether the length of the CAG or CAA repeats in each allele is less than, equal to, or greater than 29 repeats, and

- 5 determining the length of the contiguous GGN repeats in the androgen receptor gene of the subject and assessing whether the length of the GGN repeats is less than, equal to, or greater than 23 repeats, a length in at least one allele less than or greater than 29 repeats in the AIB1 gene and less than or greater than 23 repeats in the androgen receptor gene indicating an increased risk of prostate cancer in the subject,
10 wherein N is either T, G, or C.

31. The method of claim 30, wherein determining the length of the contiguous CAG or CAA repeats in the AIB1 gene alleles comprises amplifying a region of the AIB1 gene alleles comprising the CAG or CAA repeats and wherein determining the length of the contiguous GGN repeats in the androgen receptor gene
15 comprises amplifying a region of the androgen receptor gene comprising the GGN repeats.

32. The method of claim 31, wherein the amplification of the regions of the AIB1 gene and the androgen receptor gene is by PCR that produces a first and a second AIB1 PCR product and an androgen receptor PCR product.

- 20 33. The method of claim 32, further comprising analyzing the PCR products by chromatography.

34. The method of claim 33, wherein the chromatography is gel electrophoresis.

- 25 35. The method of claim 32, wherein the sequence of the PCR products is determined.

36. The method of claim 32, wherein the AIB1 PCR product is produced using a first AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:3 and a second AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:4.

- 30 37. The method of claim 36, wherein the first AIB1 primer has the sequence set forth in SEQ ID NO:1 and the second AIB1 primer has the sequence set forth in SEQ ID NO:2.

5 38. The method of claim 32, wherein the androgen receptor PCR product is produced using a first androgen receptor GGN primer that selectively hybridizes with the sequence set forth in SEQ ID NO:13 and a second androgen receptor GGN primer that selectively hybridizes with the sequence set forth in SEQ ID NO:14.

10 39. The method of claim 38, wherein the first androgen receptor GGN primer has the sequence set forth in SEQ ID NO:11 and the second androgen receptor GGN primer has the sequence set forth in SEQ ID NO:12.

15 40. The method of claim 32, wherein the AIB1 PCR product is produced using a first AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:3 and a second AIB1 primer that selectively hybridizes with the sequence set forth in SEQ ID NO:4. and wherein the androgen receptor PCR product is produced using a first androgen receptor GGN primer that selectively hybridizes with the sequence set forth in SEQ ID NO:13 and a second androgen receptor GGN primer that selectively hybridizes with the sequence set forth in SEQ ID NO:14.

20 41. The method of claim 40, wherein the first AIB1 primer has the sequence set forth in SEQ ID NO:1 and the second AIB1 primer has the sequence set forth in SEQ ID NO:2 and wherein the first androgen receptor GGN primer has the sequence set forth in SEQ ID NO:11 and the second androgen receptor GGN primer has the sequence set forth in SEQ ID NO:12.

25 42. The method of claim 30, wherein more than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and more than 23 contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

30 43. The method of claim 30, wherein more than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and less than 23 contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

 44. The method of claim 30, wherein less than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and more than 23

5 contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

45. The method of claim 30, wherein less than 29 contiguous CAG or CAA repeats in at least one allele of the AIB1 gene of the person and less than 23
10 contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

46. The method of claim 30, wherein more or less than 29 contiguous CAG or CAA repeats in both alleles of the AIB1 gene of the person and more than 23
contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

15 47. The method of claim 30, wherein more or less than 29 contiguous CAG or CAA repeats in both alleles of the AIB1 gene of the person and less than 23
contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

48. The method of claim 30, wherein more than 29 contiguous CAG or
20 CAA repeats in both alleles of the AIB1 gene of the person and more than 23
contiguous GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

49. The method of claim 30, wherein less than 29 contiguous CAG or CAA
repeats in both alleles of the AIB1 gene of the person and less than 23 contiguous
25 GGN repeats in the androgen receptor gene of the person indicates an increased risk of prostate cancer in the subject.

50. A method for assessing the risk of prostate cancer in a human subject comprising determining the length of the contiguous CAG or CAA repeats in an
AIB1 gene of the subject and assessing whether the length of the CAG repeats is less
30 than, equal to, or greater than 29 repeats, a length less than or greater than 29 repeats indicating an increased risk of prostate cancer in the subject.

51. A method for assessing the risk of prostate cancer in a human subject comprising determining the length of the contiguous CAG or CAA repeats in an

- 5 AIB1 gene of the subject and assessing whether the length of the CAG or CAA repeats is less than, equal to, or greater than 29 repeats, and determining the length of the contiguous CAG or CAA repeats in the androgen receptor gene of the subject and assessing whether the length of the CAG or CAA repeats is less than, equal to, or greater than 23 repeats, a length less than or greater than 29 repeats in the AIB1
10 allele and less than or greater than 23 repeats in the androgen receptor gene indicating an increased risk of prostate cancer in the subject.

52. A method for assessing the risk of prostate cancer in a human subject comprising determining the length of the contiguous CAG or CAA repeats in an AIB1 gene of the subject and assessing whether the length of the CAG or CAA
15 repeats is less than, equal to, or greater than 29 repeats, and determining the length of the contiguous GGN repeats in the androgen receptor gene of the subject and assessing whether the length of the GGN repeats is less than, equal to, or greater than 23 repeats, a length less than or greater than 29 repeats in the AIB1 allele and less than or greater than 23 repeats in the androgen receptor gene indicating an
20 increased risk of prostate cancer in the subject, wherein N is either T, G, or C.

53. A kit for assessing a subject's risk for acquiring prostate cancer, comprising the oligonucleotides set forth in SEQ ID Nos: 1 and 2.

54. A composition comprising the primers having the sequence set forth in SEQ ID Nos: 1 and 2.

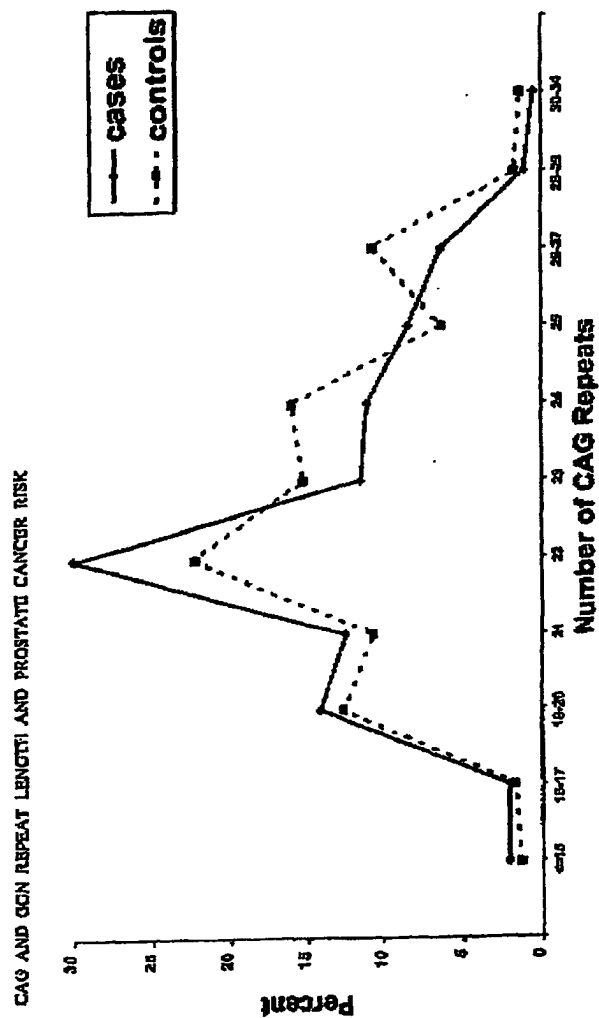


Fig. 1. Percent distribution of number of CAG repeats.

FIG. 1

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Hsing, Ann

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<210> 19

<211> 1438

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:/note =
synthetic construct

<400> 19

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Gly	Glu	Lys	Arg	Arg	Arg	Glu	Gln	Glu	Ser	Lys	Tyr	Ile	Glu	Glu	Leu	35	40	45	
Ala	Glu	Leu	Ile	Ser	Ala	Asn	Leu	Ser	Asp	Ile	Asp	Asn	Phe	Asn	Val	50	55	60	
Lys	Pro	Asp	Lys	Cys	Ala	Ile	Leu	Lys	Glu	Thr	Val	Arg	Gln	Ile	Arg	65	70	75	80
Gln	Ile	Lys	Glu	Gln	Gly	Lys	Thr	Ile	Ser	Asn	Asp	Asp	Asp	Val	Gln	85	90	95	
Lys	Ala	Asp	Val	Ser	Ser	Thr	Gly	Gln	Gly	Val	Ile	Asp	Lys	Asp	Ser	100	105	110	
Leu	Gly	Pro	Leu	Leu	Leu	Gln	Ala	Leu	Asp	Gly	Phe	Leu	Phe	Val	Val	115	120	125	
Asn	Arg	Asp	Gly	Asn	Ile	Val	Phe	Val	Ser	Glu	Asn	Val	Thr	Gln	Tyr	130	135	140	
Leu	Gln	Tyr	Lys	Gln	Glu	Asp	Leu	Val	Asn	Thr	Ser	Val	Tyr	Asn	Ile	145	150	155	160
Leu	His	Glu	Glu	Asp	Arg	Lys	Asp	Phe	Leu	Lys	Asn	Leu	Pro	Lys	Ser	165	170	175	
Thr	Val	Asn	Gly	Val	Ser	Trp	Thr	Asn	Glu	Thr	Gln	Arg	Gln	Lys	Ser	180	185	190	
His	Thr	Phe	Asn	Cys	Arg	Met	Leu	Met	Lys	Thr	Pro	His	Asp	Ile	Leu	195	200	205	
Glu	Asp	Ile	Asn	Ala	Ser	Pro	Glu	Met	Arg	Gln	Arg	Tyr	Glu	Thr	Met	210	215	220	
Gln	Cys	Phe	Ala	Leu	Ser	Gln	Pro	Arg	Ala	Met	Met	Glu	Glu	Gly	Glu	225	230	235	240
Asp	Leu	Gln	Ser	Cys	Met	Ile	Cys	Val	Ala	Arg	Arg	Ile	Thr	Thr	Gly	245	250	255	
Glu	Arg	Thr	Phe	Pro	Ser	Asn	Pro	Glu	Ser	Phe	Ile	Thr	Arg	His	Asp	260	265	270	
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Met	Arg	Pro	Gly	Phe	Glu	Asp	Ile	Ile	Arg	Arg	Cys	Ile	Gln	Arg	Phe	290	295	300	
Phe	Ser	Leu	Asn	Asp	Gly	Gln	Ser	Trp	Ser	Gln	Lys	Arg	His	Tyr	Gln	305	310	315	320
Glu	Ala	Tyr	Leu	Asn	Gly	His	Ala	Glu	Thr	Pro	Val	Tyr	Arg	Phe	Ser	325	330	335	
Leu	Ala	Asp	Gly	Thr	Ile	Val	Thr	Ala	Gln	Thr	Lys	Ser	Lys	Leu	Phe	340	345	350	
Arg	Asn	Pro	Val	Thr	Asn	Asp	Arg	His	Gly	Phe	Val	Ser	Thr	His	Phe	355	360	365	
Leu	Gln	Arg	Glu	Gln	Asn	Gly	Tyr	Arg	Pro	Asn	Pro	Asn	Pro	Val	Gly	370	375	380	
Gln	Gly	Ile	Arg	Pro	Pro	Met	Ala	Gly	Cys	Asn	Ser	Ser	Val	Gly	Gly	385	390	395	400
Met	Ser	Met	Ser	Pro	Asn	Gln	Gly	Leu	Gln	Met	Pro	Ser	Ser	Arg	Ala	405	410	415	

Tyr Gly Leu Ala Asp Pro Ser Thr Thr Gly Gln Met Ser Gly Ala Arg
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 Tyr Gly Gly Ser Ser Asn Ile Ala Ser Leu Thr Pro Gly Pro Gly Met
 435 440 445
 Gln Ser Pro Ser Ser Tyr Gln Asn Asn Asn Tyr Gly Leu Asn Met Ser
 450 455 460
 Ser Pro Pro His Gly Ser Pro Gly Leu Ala Pro Asn Gln Gln Asn Ile
 465 470 475 480
 Met Ile Ser Pro Arg Asn Arg Gly Ser Pro Lys Ile Ala Ser His Gln
 485 490 495
 Phe Ser Pro Val Ala Gly Val His Ser Pro Met Ala Ser Ser Gly Asn
 500 505 510
 Thr Gly Asn His Ser Phe Ser Ser Ser Leu Ser Ala Leu Gln Ala
 515 520 525
 Ile Ser Glu Gly Val Gly Thr Ser Leu Leu Ser Thr Leu Ser Ser Pro
 530 535 540
 Gly Pro Lys Leu Asp Asn Ser Pro Asn Met Asn Ile Thr Gln Pro Ser
 545 550 555 560
 Lys Val Ser Asn Gln Asp Ser Lys Ser Pro Leu Gly Phe Tyr Cys Asp
 565 570 575
 Gln Asn Pro Val Glu Ser Ser Met Cys Gln Ser Asn Ser Arg Asp His
 580 585 590
 Leu Ser Asp Lys Glu Ser Lys Glu Ser Ser Val Glu Gly Ala Glu Asn
 595 600 605
 Gln Arg Gly Pro Leu Glu Ser Lys Gly His Lys Lys Leu Leu Gln Leu
 610 615 620
 Leu Thr Cys Ser Ser Asp Asp Arg Gly His Ser Ser Leu Thr Asn Ser
 625 630 635 640
 Pro Leu Asp Ser Ser Cys Lys Glu Ser Ser Val Ser Val Thr Ser Pro
 645 650 655
 Ser Gly Val Ser Ser Ser Thr Ser Gly Gly Val Ser Ser Thr Ser Asn
 660 665 670
 Met His Gly Ser Leu Leu Gln Glu Lys His Arg Ile Leu His Lys Leu
 675 680 685
 Leu Gln Asn Gly Asn Ser Pro Ala Glu Val Ala Lys Ile Thr Ala Glu
 690 695 700
 Ala Thr Gly Lys Asp Thr Ser Ser Ile Thr Ser Cys Gly Asp Gly Asn
 705 710 715 720
 Val Val Lys Gln Glu Gln Leu Ser Pro Lys Lys Lys Glu Asn Asn Ala
 725 730 735
 Leu Leu Arg Tyr Leu Leu Asp Arg Asp Asp Pro Ser Asp Ala Leu Ser
 740 745 750
 Lys Glu Leu Gln Pro Gln Val Glu Gly Val Asp Asn Lys Met Ser Gln
 755 760 765
 Cys Thr Ser Ser Thr Ile Pro Ser Ser Ser Gln Glu Lys Asp Pro Lys
 770 775 780
 Ile Lys Thr Glu Thr Ser Glu Glu Gly Ser Gly Asp Leu Asp Asn Leu
 785 790 795 800
 Asp Ala Ile Leu Gly Asp Leu Thr Ser Ser Asp Phe Tyr Asn Asn Ser
 805 810 815
 Ile Ser Ser Asn Gly Ser His Leu Gly Thr Lys Gln Gln Val Phe Gln
 820 825 830
 Gly Thr Asn Ser Leu Gly Leu Lys Ser Ser Gln Ser Val Gln Ser Ile
 835 840 845
 Arg Pro Pro Tyr Asn Arg Ala Val Ser Leu Asp Ser Pro Val Ser Val
 850 855 860
 Gly Ser Ser Pro Pro Val Lys Asn Ile Ser Ala Phe Pro Met Leu Pro
 865 870 875 880
 Lys Gln Pro Met Leu Gly Gly Asn Pro Arg Met Met Asp Ser Gln Glu
 885 890 895

Asn Tyr Gly Ser Ser Met Gly Gly Pro Asn Arg Asn Val Thr Val Thr
 900 905 910
 Gln Thr Pro Ser Ser Gly Asp Trp Gly Leu Pro Asn Ser Lys Ala Gly
 915 920 925
 Arg Met Glu Pro Met Asn Ser Asn Ser Met Gly Arg Pro Gly Gly Asp
 930 935 940
 Tyr Asn Thr Ser Leu Pro Arg Pro Ala Leu Gly Gly Ser Ile Pro Thr
 945 950 955 960
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 965 970 975
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 980 985 990
 Met Gly Ala Asn Pro Tyr Gly Gln Ala Ala Ala Ser Asn Gln Leu Gly
 995 1000 1005
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 1045 1050 1055
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 1090 1095 1100
 Asp Gln Lys Ala Gly Leu Tyr Gly Gln Thr Tyr Pro Ala Gln Gly Pro
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 Met Pro Gln Ala Pro Pro Gln Gln Phe Pro Tyr Gln Pro Asn Tyr Gly
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 1330 1335 1340
 Gln His Pro Gln Ala Ala Ser Ile Tyr Gln Ser Ser Glu Met Lys Gly
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 Trp Pro Ser Gly Asn Leu Ala Arg Asn Ser Ser Phe Ser Gln Gln Gln
 1365 1370 1375

Phe Ala His Gln Gly Asn Pro Ala Val Tyr Ser Met Val His Met Asn
 1380 1385 1390
 Gly Ser Ser Gly His Met Gly Gln Met Asn Met Asn Pro Met Pro Met
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 <211> 4321
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence:/note =
 synthetic construct

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<210> 21

<211> 919

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:/note =
synthetic construct

<400> 21

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Val Ile Gln Asn Pro Gly Pro Arg His Pro Glu Ala Ala Ser Ala Ala
35          40          45
Pro Pro Gly Ala Ser Leu Leu Leu Gln Gln Gln Gln Gln Gln Gln
50          55          60
Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Glu Thr
65          70          75          80
Ser Pro Arg Gln Gln Gln Gln Gln Gln Gly Glu Asp Gly Ser Pro Gln
85          90          95
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100         105         110
Gln Pro Ser Gln Pro Gln Ser Ala Leu Glu Cys His Pro Glu Arg Gly
115         120         125

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Gln	Gln	Leu	Pro	Ala	Pro	Pro	Asp	Glu	Asp	Asp	Ser	Ala	Ala	Pro	Ser
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Thr	Leu	Ser	Leu	Leu	Gly	Pro	Thr	Phe	Pro	Gly	Leu	Ser	Ser	Cys	Ser
				165					170					175	
Ala	Asp	Leu	Lys	Asp	Ile	Leu	Ser	Glu	Ala	Ser	Thr	Met	Gln	Leu	Leu
			180					185					190		
Gln	Gln	Gln	Gln	Gln	Glu	Ala	Val	Ser	Glu	Gly	Ser	Ser	Ser	Gly	Arg
			195				200					205			
Ala	Arg	Glu	Ala	Ser	Gly	Ala	Pro	Thr	Ser	Ser	Lys	Asp	Asn	Tyr	Leu
	210					215					220				
Gly	Gly	Thr	Ser	Thr	Ile	Ser	Asp	Asn	Ala	Lys	Glu	Leu	Cys	Lys	Ala
225					230					235					240
Val	Ser	Val	Ser	Met	Gly	Leu	Gly	Val	Glu	Ala	Leu	Glu	His	Leu	Ser
				245					250					255	
Pro	Gly	Glu	Gln	Leu	Arg	Gly	Asp	Cys	Met	Tyr	Ala	Pro	Leu	Leu	Gly
				260				265					270		
Val	Pro	Pro	Ala	Val	Arg	Pro	Thr	Pro	Cys	Ala	Pro	Leu	Ala	Glu	Cys
		275					280					285			
Lys	Gly	Ser	Leu	Leu	Asp	Asp	Ser	Ala	Gly	Lys	Ser	Thr	Glu	Asp	Thr
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Ala	Glu	Tyr	Ser	Pro	Phe	Lys	Gly	Gly	Tyr	Thr	Lys	Gly	Leu	Glu	Gly
305					310					315					320
Glu	Ser	Leu	Gly	Cys	Ser	Gly	Ser	Ala	Ala	Ala	Gly	Ser	Ser	Gly	Thr
				325					330					335	
Leu	Glu	Leu	Pro	Ser	Thr	Leu	Ser	Leu	Tyr	Lys	Ser	Gly	Ala	Leu	Asp
			340					345					350		
Glu	Ala	Ala	Ala	Tyr	Gln	Ser	Arg	Asp	Tyr	Tyr	Asn	Phe	Pro	Leu	Ala
		355					360					365			
Leu	Ala	Gly	Pro	Pro	Pro	Pro	Pro	Pro	Pro	Pro	His	Pro	His	Ala	Arg
	370						375					380			
Ile	Lys	Leu	Glu	Asn	Pro	Leu	Asp	Tyr	Gly	Ser	Ala	Trp	Ala	Ala	Ala
385					390					395					400
Ala	Ala	Gln	Cys	Arg	Tyr	Gly	Asp	Leu	Ala	Ser	Leu	His	Gly	Ala	Gly
				405					410					415	
Ala	Ala	Gly	Pro	Gly	Ser	Gly	Ser	Pro	Ser	Ala	Ala	Ala	Ser	Ser	Ser
			420					425					430		
Trp	His	Thr	Leu	Phe	Thr	Ala	Glu	Glu	Gly	Gln	Leu	Tyr	Gly	Pro	Cys
		435					440					445			
Gly	Gly	Gly	Gly	Gly	Gly	Gly	Gly	Gly	Gly	Gly	Gly	Gly	Gly	Gly	Gly
	450					455					460				
Gly	Gly	Gly	Gly	Gly	Gly	Gly	Gly	Glu	Ala	Gly	Ala	Val	Ala	Pro	Tyr
465					470					475					480
Gly	Tyr	Thr	Arg	Pro	Pro	Gln	Gly	Leu	Ala	Gly	Gln	Glu	Ser	Asp	Phe
				485				490						495	
Thr	Ala	Pro	Asp	Val	Trp	Tyr	Pro	Gly	Gly	Met	Val	Ser	Arg	Val	Pro
			500					505					510		
Tyr	Pro	Ser	Pro	Thr	Cys	Val	Lys	Ser	Glu	Met	Gly	Pro	Trp	Met	Asp
		515					520					525			
Ser	Tyr	Ser	Gly	Pro	Tyr	Gly	Asp	Met	Arg	Leu	Glu	Thr	Ala	Arg	Asp
	530					535					540				
His	Val	Leu	Pro	Ile	Asp	Tyr	Tyr	Phe	Pro	Pro	Gln	Lys	Thr	Cys	Leu
545					550					555					560
Ile	Cys	Gly	Asp	Glu	Ala	Ser	Gly	Cys	His	Tyr	Gly	Ala	Leu	Thr	Cys
				565					570					575	
Gly	Ser	Cys	Lys	Val	Phe	Phe	Lys	Arg	Ala	Ala	Glu	Gly	Lys	Gln	Lys
			580					585					590		
Tyr	Leu	Cys	Ala	Ser	Arg	Asn	Asp	Cys	Thr	Ile	Asp	Lys	Phe	Arg	Arg
		595					600					605			

Lys Asn Cys Pro Ser Cys Arg Leu Arg Lys Cys Tyr Glu Ala Gly Met
 610 615 620
 Thr Leu Gly Ala Arg Lys Leu Lys Lys Leu Gly Asn Leu Lys Leu Gln
 625 630 635 640
 Glu Glu Gly Glu Ala Ser Ser Thr Thr Ser Pro Thr Glu Glu Thr Thr
 645 650 655
 Gln Lys Leu Thr Val Ser His Ile Glu Gly Tyr Glu Cys Gln Pro Ile
 660 665 670
 Phe Leu Asn Val Leu Glu Ala Ile Glu Pro Gly Val Val Cys Ala Gly
 675 680 685
 His Asp Asn Asn Gln Pro Asp Ser Phe Ala Ala Leu Leu Ser Ser Leu
 690 695 700
 Asn Glu Leu Gly Glu Arg Gln Leu Val His Val Val Lys Trp Ala Lys
 705 710 715 720
 Ala Leu Pro Gly Phe Arg Asn Leu His Val Asp Asp Gln Met Ala Val
 725 730 735
 Ile Gln Tyr Ser Trp Met Gly Leu Met Val Phe Ala Met Gly Trp Arg
 740 745 750
 Ser Phe Thr Asn Val Asn Ser Arg Met Leu Tyr Phe Ala Pro Asp Leu
 755 760 765
 Val Phe Asn Glu Tyr Arg Met His Lys Ser Arg Met Tyr Ser Gln Cys
 770 775 780
 Val Arg Met Arg His Leu Ser Gln Glu Phe Gly Trp Leu Gln Ile Thr
 785 790 795 800
 Pro Gln Glu Phe Leu Cys Met Lys Ala Leu Leu Leu Phe Ser Ile Ile
 805 810 815
 Pro Val Asp Gly Leu Lys Asn Gln Lys Phe Phe Asp Glu Leu Arg Met
 820 825 830
 Asn Tyr Ile Lys Glu Leu Asp Arg Ile Ile Ala Cys Lys Arg Lys Asn
 835 840 845
 Pro Thr Ser Cys Ser Arg Arg Phe Tyr Gln Leu Thr Lys Leu Leu Asp
 850 855 860
 Ser Val Gln Pro Ile Ala Arg Glu Leu His Gln Phe Thr Phe Asp Leu
 865 870 875 880
 Leu Ile Lys Ser His Met Val Ser Val Asp Phe Pro Glu Met Met Ala
 885 890 895
 Glu Ile Ile Ser Val Gln Val Pro Lys Ile Leu Ser Gly Lys Val Lys
 900 905 910
 Pro Ile Tyr Phe His Thr Gln
 915

INTERNATIONAL SEARCH REPORT

(PCT Article 18 and Rules 43 and 44)

Applicant's or agent's file reference CG/12326.19	FOR FURTHER ACTION see Notification of Transmittal of International Search Report (Form PCT/ISA/220) as well as, where applicable, Item 5 below.	
International application No. PCT/CA 99/ 00852	International filing date (day/month/year) 15/09/1999	(Earliest) Priority Date (day/month/year) 15/09/1998
Applicant SIGNALGENE INC. et al.		

This International Search Report has been prepared by this International Searching Authority and is transmitted to the applicant according to Article 18. A copy is being transmitted to the International Bureau.

This International Search Report consists of a total of 3 sheets.

☒ It is also accompanied by a copy of each prior art document cited in this report.

1. Basis of the report

a. With regard to the language, the international search was carried out on the basis of the international application in the language in which it was filed, unless otherwise indicated under this item.

☐ the international search was carried out on the basis of a translation of the international application furnished to this Authority (Rule 23.1(b)).

b. With regard to any nucleotide and/or amino acid sequence disclosed in the international application, the international search was carried out on the basis of the sequence listing :

☐ contained in the international application in written form.

☐ filed together with the international application in computer readable form.

☒ furnished subsequently to this Authority in written form.

☒ furnished subsequently to this Authority in computer readable form.

☒ the statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.

☒ the statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished

2. ☐ Certain claims were found unsearchable (See Box I).

3. ☐ Unity of invention is lacking (see Box II).

4. With regard to the title,

☒ the text is approved as submitted by the applicant.

☐ the text has been established by this Authority to read as follows:

5. With regard to the abstract,

☒ the text is approved as submitted by the applicant.

☐ the text has been established, according to Rule 38.2(b), by this Authority as it appears in Box III. The applicant may, within one month from the date of mailing of this international search report, submit comments to this Authority.

6. The figure of the drawings to be published with the abstract is Figure No.

☐ as suggested by the applicant.

☐ because the applicant failed to suggest a figure.

☐ because this figure better characterizes the invention.

☐ None of the figures.